Phytochemical Screening And Antimicrobial Study Of The Different Leaf Extracts Of *Alocasia sanderiana* Bull., An Endemic Philippine Plant

Romeo C. Ongpoy Jr.

Abstract: The objective of this study is to investigate the phytochemical contents and evaluate the antimicrobial property of *Alocasia sanderiana* Bull. against a large number of pathogens. To do this, *Alocasia sanderiana* Bull. was screened for qualitative phytochemical tests including thin layer chromatography. Aside from the crude extract from the Rotary evaporator, three fractions from the plant were prepared using methanol, dichloromethane (DCM) and hexane. The 4 solvent extracts were then evaluated for antimicrobial activity using disc diffusion method on 18 strains of organisms. About this study, it was found out that triterpenes, tannins and saponins are present during phytochemical screening. Zones of inhibitions during the antimicrobial tests were observed but did not reach the desired zone for antimicrobial activity. The DCM fraction produced 4 mm zone against *Proteus mirabilis*, 3 mm for *Pseudomonas aeruginosa*, 1 mm for *Pectobacterium carotovorum* and 1 mm for *Candida albicans*. The methanol fraction also produced a 1 mm zone against *Pseudomonas aeruginosa*. The results show that *Alocasia sanderiana* Bull. leaf extracts contain polyphenolic compounds but this study shows that it exhibits non-active antimicrobial activity against the 18 strains that it was tested and may not be utilized as a potential antimicrobial drug for the said strains.

Index Terms: Alocasia sanderiana Bull., Antimicrobial, Phytochemical screening

1 INTRODUCTION

Alocasia sanderiana Bull. is an endemic Philippine plant, although there has been studies on other Alocasia spp., there hasn't been any for the subject plant. Alocasia spp. has been used for economic purposes mainly as food¹ from its leaf to the whole plant depending on the specie but for Alocasia sanderiana Bull., it serves as an ornamental in Filipino houses. This study would like to find out if it may be used as a potential spp. There are evidently less studies undertaken for this genus and none for Alocasia sanderiana Bull. but should a medicinal or pharmaceutical use be discovered for this plant, it may become more than just an ornamental in the Filipino houses. Medicinal studies Alocasia include on spp. the hepatoprotective effect^{5,6} of Alocasia indica Linn. which may be due to an antioxidant property that may be present as in Alocasia macrorrhiza Linn.

A. macrorrhiza has also been recorded for its folkloric use in Malaysia for cough and toothache⁸ as well as in parasitic infestations⁹. On the other hand, Alocasia odora (Lindl.) K. Koch burned leaves have also been used in folkloric medicine as a liniment against small pox.¹⁰ Another interesting medicinal use of the Alocasia spp. particularly Alocasia Cucullata (Lour) Schott. is its use against snake bites¹¹ where it is believed to be an anti-venom^{12,13,14} but there are no scientific studies to prove this medicinal property. Alocasia spp. is not all about medicinal properties; in fact, there are numerous literatures that classify it as poison^{15,16} with one literature citing it as part of the 55 cases of herb-induced poisoning in a hospital based study from 1995 to 2007¹⁷ that may cause neurotoxicity^{18,1} which may be due to its calcium oxalate content²⁰ and other chemical irritants that also cause local pain, swelling, blistering of skin and mucous membranes²¹ as well as dermatitis^{22,23}. It may also cause excessive salivation and oral irritation²⁴ contrary to its use as food. It is also recorded that Alocasia spp. causes toxic reaction in pets usually birds²⁵. This conflicting literature calls for a need to further study Alocasia spp. including Alocasia sanderiana Bull. because there is no clear information for its safety, there is no available scientific study for its antimicrobial property and there is even no study for the specie itself. Alocasia sanderiana Bull. may cause irritation and even poisoning owing to its calcium oxalate content that was observed microscopically by the researcher upon histological examination of the plant leaf but calcium oxalates are easily removed after heating²⁶ which may enable clinical trials for antibacterial property after this pre-clinical study. The purpose of this study is to confirm its indirect use as antimicrobial which is still not backed by scientific data up to the present day which may be due to its phytochemical constituents particularly the polyphenols after removal of the calcium oxalate as part of the plant extract concentration through the use of a rotary evaporator.

[•] Romeo C. Ongpoy, Jr. MSc RPh CPS is an Associate Professor at the College of Pharmacy, De La Salle Health Science Institute / School of Health Science Professions, St. Dominic College of Asia. He is also a Regulatory Affairs Associate at the Regulatory, Quality and Clinical Department, PerkinElmer Instruments (Philippines) Corporation and currently pursuing a PhD Pharmacy degree in Centro Escolar University, Mendiola in the Philippines.

Telephone number: +63 2 9167205641and E-mail address: rongpoy@gmail.com

2 METHODOLOGY

2.1 Plant Materials

The plant material was collected in December 2012 at Salay, Misamis Oriental where it was asexually cultivated for research purposes in a loamy soil. The samples were washed, air-dried for 30 days in a clean well-ventilated area and packed in a polyethylene bag for transport in Manila. The botanical identification of species was carried out by the scientists at the Botany Division of the Philippine National Museum.

2.2 Materials

Methanol, Hexane and DCM were all obtained from RCI Labscan Limited through their local distributor, Belman Laboratories. Mueller Hinton agar and Sabouraud Dextrose agar were obtained from Sigma-Aldrich Chemicals through the same local distributor as well as the 6mm Antibiotic Assay Disk from GE.

 TABLE 1

 PHYTOCHEMICAL SCREENING OF ALOCASIA SANDERIANA BULL.

 LEAF EXTRACT

Phytoconstituents	Test Tube Reaction		Thin Layer Chromatography				
	Method	Result	Method	Result			
Glycosides		Positive					
Tannins		Positive (non-	Ferricyanide-	Positive			
		hydrolysable)	ferric chloride				
Alkaloids		Negative	Dragendorff's	Negative			
Organic Acids		Negative					
Sterols and		Positive	Vanillin-	Positive			
Triterpenes			sulfuric acid				
Saponins		Positive					
Flavonoids		Negative					
Starch		Positive					
Albuminoids		Positive					
Sugars			α-Naphthol-	Positive			
			sulfuric acid				
	Molisch's	Positive					
	lodine	Positive					
	Anthrone	Positive	Borntrager	Positive			
	Barfoed's	Negative					
	Seliwanoff;s	Negative					
	Benedicts's	Positive					

2.3 Extraction of the Plant Material

The leaves of *A. sanderiana* Bull were cut to pieces and milled to fine powder then percolated with methanol at 1:10 ratio²⁷ for 48 hours. The percolated material was then filtered and the filtrate underwent concentration through Rotary Evaporator. The concentrated sample was subjected to fractionation using Vacuum Liquid Chromatography using 3 solvents added in the following order: methanol>DCM>hexane. The fractions were then stored at 4°C when not in use. The antimicrobial assay was performed using the fractions while the crude drug product from the Rotary Evaporator was used for the Phytochemical analysis.

2.4 Phytochemical Screening

Phytochemical screening for major phytoconstituents of the plant extracts was undertaken using standard qualitative methods as described by various authors^{28,29}. The plant extracts were screened of biologically active compounds like glycosides, tannins, alkaloids, organic acids, sterol and triterpenes, saponins, flavonoids, starch, albuminoids, sugars. Thin layer chromatography was also used to confirm the

previous test tube methods where it was found out that 1 part hexane and 9 parts ethyl acetate may be used as the solvent mixture but pure ethyl acetate was eventually used due to the better separation of compounds.

 TABLE 2

 ANTIMICROBIAL ACTIVITY OF THE DIFFERENT LEAF EXTRACTS OF

 ALOCASIA SANDERIANA BULL.

	Zone of Inhibition (mm)					
Microorganisms	Crude Extract	Methanol	DCM Fraction	Hexane		
-		Fraction		Fraction		
Gram +	0	0	0	0		
E. faecalis	0	0	0	0		
S. pyogenes	0	0	0	0		
S. pneumoniae	0	0	0	0		
B. subtilis	0	0	0	0		
S. aureus	0	0	0	0		
MRSA	0	0	0	0		
S. epidermidis	0	0	0	0		
B. cereus	0	0	0	0		
Gram -	0	0	0	0		
P. mirabilis	0	0	4 mm	0		
E. coli	0	0	0	0		
S. marcescens	0	0	0	0		
E. aerogenes	0	0	0	0		
P. carotovorum	0	0	1 mm	0		
S. typhimurium	0	0	0	0		
P. aeruginosa	0	1 mm	3 mm	0		
S. cerevisae	0	0	0	0		
Fungus	0	0	0	0		
C. albicans	0	0	1 mm	0		
S. cerevisiae	Ö	Ö	0	0		
A. niger	0	Ö	0	0		

2.5 Antimicrobial Assay 2.5.1 Test Organisms

The test microorganisms used in this study included 18 strains all obtained from the University of Santo Tomas - Tomas Aguinas Research Center (TARC). The Gram positive organisms were Enterococcus faecalis, Streptocuccus pyogenes (ATCC 19615), Streptococcus pneumoniae (ATCC 49619), Bacillus subtilis (UST CMS 1011), Staphylococcus aureus (ATCC 29213), Staphylococcus aureus MRSA (ATCC 43300), Staphylococcus epidermidis (ATCC 12228), Bacillus cereus (UST CMS 1009). The Gram negative organisms were Proteus mirabilis (UST CMS 1070), Escherichia coli (ATCC 25922), Serratia marcescens (UST CMS 1095), Enterobacter aerogenes (UST CMS 1021) Pectobacterium carotovorum, Salmonella typhimurum, Pseudomonas aeruginosa (ATCC 27853). The fungal specimens were Candida albicans (UST CMS 1201), Saccharomyces cerevisiae (UST CMS 1211), Aspergillus niger.

2.5.2 Inoculum Preparation

The microbial suspensions were standardized from previously conserved strains at TARC and inoculated at Mueller-Hinton broth for bacteria and Sabouraud Dextrose broth for fungi at 37°C. After 24 hours of incubation, suspensions were diluted. Inocula were set to 0.5 McFarland equivalent to an optical density from 0,08 to 0.13 at 625 nm wavelength, which corresponds to 108 CFU/mL^{30,31}.

2.5.3 Disc Diffusion Method

Disposable Petri dish (9 cm) were prepared with 20 mL of a base layer of molten Mueller Hinton agar for bacteria and Sabouraud Dextrose agar for fungi. Each Petri dish was inoculated with 15 uL of bacterial suspension or fungal suspension equivalent to 10⁶ CFU/mL³². After drying in a hood,



6mm diameter discs were each added separately with 20 uL *Alocasia sanderiana* Bull. fractions an crude extract using a micropipette. The plates were then incubated for 24 hours at 37°C for bacteria³³ and 48 hours at 37°C for fungi. The diameters of the zones were evaluated in millimeters and fractions inducing inhibition zones at least 8 mm³⁴ around the disc were considered antimicrobial. All tests were performed in triplicate³⁵.

3 RESULTS

3.1 Phytochemical Screening

The phytochemical screening of the crude extracts of *Alocasia* sanderiana Bull. using test tube methods showed that glycosides, non-hydrolysable tannins, sterols and triterpenes, saponins starch, albuminoids and most sugars are present while colorimetric method using TLC futher confirmed the presence of tannins, triterpenes and sterols and some sugars, it also showed the presence of anthrone specifically (Table 1).

3.2 Antimicrobial Activity

The antibacterial activity of *Alocasia sanderiana* Bull. leaves is non-active using the criteria of at least 8 mm zone of inhibition for antimicrobial activity. There are zones though that may be seen below the 8 mm criteria, the DCM fraction showed 4 mm zones for *P. mirabilis*, 3 mm zones for *P. aeruginosa* and 1 mm zones for *P. carotovorum* and *C. albicans*. The methanol fraction also showed 1 mm zone of inhibition against *P. aeruginosa* (Table 2).

4 **DISCUSSION**

Although Alocasia sanderiana Bull. leaf extracts contain polyphenolic compounds this study shows that it has no antimicrobial activity against the 18 microorganisms that it was tested and may not be utilized as a potential antimicrobial drug. There are small zones of inhibitions though that were observed in some microorganisms but it was not enough for the minimum of 8 mm zone applied to this study, this zone however may be due to the presence of protease inhibitors discovered in other Alocasia spp.36 that may be present in Alocasia sanderiana Bull. but there may not be enough present in the plant to cause an antimicrobial property. There are also other compounds like trypsin inhibitors^{37,38} and lectins^{39,40} that are present in other *Alocasia* spp. where its role is not yet well studied in Alocasia spp. that may affect its antimicrobial property. Therefore more bioactive components need be discovered and may be possible through the use of instrumentation like Gas Chromatography⁴¹ like in the recent case of Alocasia indica (Lour.) Spach. It is also important to note that there are different interpretations in the zone of inhibitions, this study focused on the more commonly used minimum zone of inhibition for an antibacterial effect but some studies are utilizing at least 3 mm as weak antibacterial activity. Future studies may focus on its toxicity data to be able to further study its medicinal potential like in coagulation due to the mentioned trypsin inhibitors present in other species and in viral studies owing to the protease inhibitors in its other species as well. One of the limitations of this study is that it only focused on the leaves, other plant organs like roots (tubers) may also be studied in the future as there are folkloric utilizations of the organ.

Acknowledgment

The author gratefully acknowledges the assistance of Ms. Chiristina Tan RCh for sharing her expertise. This research is also being supported by the Philippine Council of Health Research and Development

References

- [1] Deb, S., Arunachalam, A., Das, AK. Indigenous Knowledge of Nyishi Tribes on Traditional Agroforestry Systems. Indian Journal of Traditional Knowledge 2009; 8(1): 41-46.
- [2] Tien, NQ., Ngoc, PH., Minh, PH., Kiem, PV., Minh, CV., Kim, YH. New Ceramide from Alocasia macrorrhiza. Archives of Pharmacal Research 2004; 27(10): 1020-1022.
- [3] Chuakul, W., Saralamp, P. Survey on Medicinal Plants Used in Khok Phayuung Village, Kaapchoeng District, Surin Province, Thailand. Journal of National Research Council Thailand 2002; 34(1).
- [4] Wang, Y., Yin, J., Xu, Z. Alocasia hypnosa (Araceae), A New Species from Yunan, China. Annals of Botanical Fennici October 2005; 42: 395-398.
- [5] Mulla, W., Salunkhe, V., Bhise, S. Hepatoprotective Activity of Hydroalcoholic Extract of Leaves of Alocasia indica (Linn.) IndianJournal of Experimental Biology 2009; 47: 816-821.
- [6] Kotoky, J., Das, PN. Medicinal Plants Used for Liver Diseases in Some Parts of Kamrup District of Assam, A North Eastern State of India. Fitoterapia July 2008; 79(5): 384-387.
- [7] Choi, E., Hwang, JK. Screening of Indonesian Medicinal Plants for Inhibitor Activity on Nitric Oxide Production of RAW264.7 Cells and Antioxidant Activity. Fitoterapia March 2005; 76(2): 194-203.
- [8] Sulaiman, B., Mansor, M. Medicinal Aroids Conservation: A Case Study of Floral Garden, School of Biological Sciences, Universiti Sains, Malaysia. Proceedings of the 4th IMT-GT UNITET Conference 2002.
- [9] Wilcox, ML., Bodeker, G. (2004). Herbal Remedies for Malaria: An Overview of Clinical Studies. UK: Harwood Press.
- [10] Inta, A. et al. A Comparative Study on Medicinal Plants Used in Akha's Traditional Medicine in China and Thailand, Cultural Coherence or Ecological Divergence?. Journal of Ethnopharmacology 28 March 2008; 116(3): 508-517.
- [11] Otero, R., Nunez, V., Barona, J., Fonnegra, R., Jimenez, SL., Osorio, RG., Saldarriaga, M., Diaz, A. Snakebites and Ethnobotany in the Northwest Region of Colombia: Part III: Neutralization of the Haemorrhagic Effect of Bothrops atrox Venom.

Journal of Ethnopharmacology November 2000; 73(1-2): 233-241.

- [12] Houghton, P. In vitro Testing of some West African and Indian Plants Used to Treat Snakebites. European Society of Ethnopharmacology 1993.
- [13] Martz, W. Plants with a Reputation Against Snakebite. Toxicon 1992; 30: 1131-1142.
- [14] Soraes, AM., Ticli, FK et al. Medicinal Plants with Inhibitory Properties Against Snake Venoms. Current Medicinal Chemistry October 2005; 12(22): 2625-2641.
- [15] Carles, M, Cheung, MKL, Moganti, S., Dong, TTX., Tsim, KW., Ip, NY., Sucher, NJ. A DNA Microarray for the Authentication of Toxic Traditional Chinese Medicinal Plants. Planta Medica 2005; 71: 580-584.
- [16] Goonasekera, CDA., Vasanthathilake, VWJK., Ratnatunga, N., Seneviratne, CAS. Is Nai Habarala (Alocasia cucullata) A Poisonous Plant?. Toxicon June 1993; 31(6): 813-816.
- [17] Colombo, ML., Assisi, F., Puppa, TD., Moro, P., Sesana, FM., Bissoli, M., Borghini, R., Perego, S., Galasso, G., Banfi, E., Davanzo, F. Exposures and Intoxications After Herb-induced Poisoning: A Retrospective Hospital-based Study. Journal of Pharmaceutical Sciences and Research 2009; 2(2): 123-136.
- [18] Moon, JM., Lee, BK., Chun, BJ. Toxicities of Raw Alocasia odora. Human and Experimental Toxicology October 2011; 30(10): 1720-1723.
- [19] Chan, TY., Chan, LY., Tam, LS., Critchley, JA. Neurotoxicity following the Ingestion of a Chinese Medicinal Plant, Alocasia macrorrhiza. Human and Experimental Toxicology 1995; 14(9): 727-728.
- [20] Tagwireyi, D., Ball, DE. The Management of Elephant's Ear Poisoning. Human and Experimental Toxicology April 2001; 20(4): 189-192.
- [21] Moro, PA., Assisi, F., Cassetti, F., Bissoli, M., Borghini, R., Davanzo, F., Della Puppa, T., Dimasi, V., Ferruzzi, M., Giarratana, T., Travaglia, A. Toxicological Hazards of Natural Environment: Clinical Reports from Poison Control Centre of Milan. Urban Forestry and Urban Greening 2009; 8(3): 179-186.
- [22] Botha, CJ., Penrith, ML. Poisonous Plants of Veterinary and Human Importance in Southern Africa. Journal of Ethnopharmacology 28 October 2008; 119(3): 549-558.
- [23] Wagstaff, DJ. (2008). International Poisonous Plants Checklist: An Evidence-Based Reference. USA: Taylor and Francis Group.

- [24] Peterson, ME. Talcott, PA. (2013). Small Animal Toxicology, 3rd edition. USA: Saunders.
- [25] Schulte, MS., Rupley, AE. The Veterinary Clinics of North America: Exotic Animal Practice. Avian Care and Husbandry 2004; 7(2): 315-350.
- [26] Catherwood, DJ. Et al. Oxalate Content of Cormels of Japanese Taro and the Effect of Cooking. Journal of Food Composition and Analysis 2007; 20: 147-151.
- [27] Ansel, H. (2010). Pharmaceutical Calculations, 13th edition. USA: Lippincott Williams & Wilkins. 319-322.
- [28] Austria, Z., Lontoc, B., Perez, T. (2007). A Research Compedindum of 100 Wild Plants. Manila: Centro Escolar University. 207-224.
- [29] Guevara G. (2005). A Guidebook to Plant Screening: Phytochemical and Biological. Manila: University of Santo Tomas. 24-57, 103-110, 120-134.
- [30] Moura-Costa, GF., Nocchi, SR., Ceole, LF., de Mello, JCP., Nakamura, CV., Filho, BPD., Temponi, LG., Ueda-Nakamura, T. Antimicrobial Activity of Plants Used as Medicinals on an Indigenous Reserve in Rio das Cobras, Parana, Brazil. Journal of Ethnopharmacology 2012; 143: 631-638.
- [31] Benbelaid, F., Khadir, A., Abdoune, MA., Bendahuo, M. Phytochemical Screening and in vitro Antimicrobial Activity of Thymus lanceolatus Desf. From Algeria. Asian Pacific Journal of Tropical Disease 2013; 3(6): 454-459.
- [32] Konate, K., Yomalan, K., Sytar, O., Brestic, M. Antidiarrheal and Antimicrobial Profiles Extracts of the Leaves from Trichilia emetic Vahl. (Meliaceae). Asian Pacific Journal of Tropical Biomedicine 2015; 5(3): 242-248.
- [33] Khatoon, M., Khatun, H., Islam, E., Parvin, S. Analgesic, Antibacterial and Central Nervous System Depressant Activities of Albizia procera Leaves. Asian Pacific Journal of Tropical Biomedicine 2014; 4(4): 279-285.
- [34] Bouabdelli, F., Djelloul, A., Kaid-Omar, Z., Semmoud, A., Addou, A. Antimicrobial Activity of 22 Plants Used in Urolithiasis Medicine in Western Algeria. Asian Pacific Journal of Tropical Disease 2012; 8530-8535.
- [35] Van Vuuren, SF., Naidoo, D. An Antimicrobial Investigation of Plants Used Traditionally in Southern Africa to Treat Sexually Transmitted Infections. Journal of Ethnopharmacology 2010; 130: 552-558.
- [36] Polya, GM. Protein and Non-Protein Protease Inhibitors from Plants. Studies in Natural Products Chemistry 2003; 29: 567-641.

- [37] Wang, HX., Ng, TB. Alocasin, an Anti-fungal Protein From Rhizomes of the Giant Taro (Alocasia macrorrhiza). Protein Expression and Purification March 2003; 28(1): 9-14.
- [38] Argall, ME., Bradburry, JH., Shaw, DC. Amino Acid Sequence of a Trypsin/Chemotrypsin Inhibitor From Giant Taro (Alocasia macrorrhiza). Biochimica et Biophysica Acta (BBA) – Protein Structure and Molecular Enzymology 1994; 1204(2): 189-194.
- [39] Dhuna, V., Sharma, K., Singh, J., Kamboj, SS. Purification and Characterization of Monocot Lectin from Tubers of Alocasia indica Having in vitro Antiproliferative Effect on Human Cancer Cell Lines. Cancer Research 2009; 69: 2674.
- [40] Kamboj, SS., Shangary, S., Singh, J., Kamboj, JJ., Sandhu, RS. New Lymphocyte Stimulating Monocot Lectins from Family Araceae. Immunological Investigations: A Journal of Molecular and Cellular Immunology 2005; 24(5): 845-855.
- [41] Basu, S., Das, M., Sen, A., Choudhury, UR., Datta, G. Analysis of Complete Nutritional Profile and Identification of Bioactive Components Present in Alocasia indica Tuber Cultivated in Howrah District of West Bengal, India. Asian Pacific Journal of Tropical Medicine 2014; 7(suppl 1): S527-S533.

