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REVIEW ON PHYTOPHARMACOLOGICAL ACTIVITIES OF ANDROGRAPHIS PANICULATA (BURM.F) NEES.

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ABSTRACT

Medicinal plants are in prevalence from the origin of the earth which has been efficiently used by ancient people as a cure for various ailments. The plant extracts are identified and act as an important source of active ingredient and secondary metabolite products such as alkaloid, terpenoids, which is used in curing diseases, for drug production and perpetuating good health by both the traditional and orthodox medical practitioners. The current aim of this review is to accumulate the morphological and pharmacological applications of Andrographis paniculata as a multipurpose drug showing efficient activity in curing various diseases. The plant is highly useful in curing various health ailments of human beings especially in viral diseases such as respiratory problems. HIV. Liver damage etc. The phytochemical tests revealed the presence of glycosides, saponins, tannins and alkaloids, but not of anthraguinones. Thus the plant has an important compound named as Andrographolide, a diterpenoid lactone having a diversity of pharmacological effects. The overdosage of the plant leads to some side effects like nausea, vomiting and loss of appetite Therefore, researchers have to perform various formulation for A. paniculata and develop a new drug molecules. The plant is to be widely cultivated and farmers should be encouraged in cultivating the medicinal plant which has multifarious therapeutic uses. The plant is considered as a safe, highly important medicinal plant for mankind.

KEY WORDS: Andrographis paniculata, Biological Activities, Molecular characters, Elemental analysis, Anti HIV.

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INTRODUCTION

Medicinal and aromatic plants are spread throughout the forest areas of South Asian places both in the tropical and subtropical belts. India has a rich diversity of plants having the medicinal values. Some plants are ubiguitously found in extreme environmental conditions, where plants grow very gradually and cannot live in any other environment. While some other plants are widely distributed and it can acclimatize more easily to various other ecological conditions. The primary metabolites such as proteins, phenols, sugars, starch and lipids which are useful in flavoring, fragrances, insecticides, sweeteners and natural dyes, are essentially required for the growth of plants.¹ The medicinal plants are also rich in secondary metabolites which have been used as a drug in pharmaceutical industry which includes alkaloids, glycosides, amines, insecticides, steroids, phenols, saponins, tannins, terpenoids, flavonoids etc.²The significant source for plants is the antioxidant activity which produces secondary metabolites. The antioxidant properties present in natural substances (food) for medicinal and preservative purposes³. An antioxidant terminates the chain reaction of a molecule that damages the cell by removing free radical intermediates present in it and then the oxidation reactions are inhibited by reducing the stress responsible for many degenerative disorders⁴⁻⁵. Natural products of higher plants may possess a new source of antimicrobial agents⁶⁻⁷. Thy infectious diseases are treated by the medicinal plants which mitigate many of the side effects which are associated with synthetic antimicrobials⁸. Cancer is a major dreadful disease caused by abnormal or uncontrolled cell division. About 6 million new occurrences of cancer are reported yearly worldwide. In the discovery of drug and its development; especially its activity against cancer and other infectious diseases⁹ were natural products have played a significant role are prone mostly in low and middle income countries. The risk factors of cancer reported by WHO, noted that the use of tobacco, alcohol, low fruit and vegetable intake, and chronic infections are caused by hepatitis B virus (HBV), hepatitis C virus (HCV) and some types of human papilloma virus (HPV) are prevalent in the above mentioned countries¹⁰.Mankind is most commonly affected by the widespread of vicious diseases like Malaria. The causative agent of the disease is caused by Plasmodium (Protozoa) of which four species *P. falciparam, P.vivax, P.ovale* and *P.malariae* are contagious to humans¹¹. The disease is epidemic in tropical and sub-tropical areas of the world, especially because of the pitiless rise in the resistance of *P.falciparum* to commonly used antimalarial drugs like chloroquine, mefloquine, mepacrine, pyrimethamine, primaquine and sulphadoxin¹²⁻¹³. Viruses and bacteria present in the environment may cause upper respiratory infection. The occurrence of infection in nose, throat,

sinuses, and ears is termed as upper respiratory infection. eg: common cold¹⁴⁻¹⁵. Three ingredients deoxyandrographolide, Andrographolide and neoandrographolide effective in reducing are inflammation¹⁶. Dermatophytosis is a fungal infection is one of the major health problems in tropical countries. The retroviridae family encompasses a group of viruses in which the replicative life cycle requires reverse transcription of the viral ribonucleic acid (RNA) genome into double stranded deoxyribonucleic acid (DNA). Retroviridae initially were identified in a variety of animal species in association with neoplasms such as leukemia and lymphoma; it subsequently was discovered that endogenous retroviral sequences are abundant in humans, comprising at least 1% of the human genome and demonstrating Mendelian inheritance patterns

¹⁹.The human immunodeficiency virus, or HIV, is the virus that causes HIV infection. The virus attacks and destroys the infection-fighting CD4 cells of the body's immune system. Loss of CD4 cells makes it difficult for the immune system to fight against the infections is transmitted (spread) through the blood, semen, genital fluids, or breast milk of a person infected with HIV. The main reason for the transmission of HIV to a healthy person is by unprotected sex or sharing drug injection equipment such as needles and syringes. A diagnosis of HIV is based on the first test that can be either an HIV antibody test (using blood, urine, or fluids from the mouth) or a plasma HIV RNA test (using blood). The second test always using blood is a different type of antibody test called a Western blot test. A positive Western blot test confirms that a person has HIV. Andrographis paniculata Nees belongs to the family of Acanthaceae commonly known as "Kalmegh" or "King of bitters". It is a perennial herb which is about 32 different species present in the genus Andrographis. Around 175 BC the plant Andrographis paniculata was recommended in Charaka samhita for the treatment of jaundice along with other plants in polyherbal preparations. In India, it is cultivated during rainy phase of summer season (kharif crop). The amount of organic matter present in the soil is best suitable for commercial cultivation of the crop.²⁰In world, it is widely cultivated in China, Indonesia, Hong Kong, Philipines, the East and Mauritius ²⁰⁻²¹ and West Indies such as Jamaica, Barbados, and Bahamas¹¹⁵Thailand, South Asia, South Africa, Java, Malaysia, Brunei, Singapore, India, Pakistan and Srilanka¹¹⁰⁻¹¹¹. Cambodia, Caribbean islands, Laos, Myanmar, and Vietnam¹¹²⁻¹¹³. This plant is also found in different phytogeographical and edaphic zones of China, America, West Indies, and Christmas Island¹¹⁴. In India, it is cultivated from Uttar Pradesh, Himachal Pradesh, Assam, Madhya Pradesh, Tamil Nadu, Karnataka and Kerala. In Tamil Nadu, it is cultivated in Tanjore, salem, Erode, Villupuram, Tiruchengode and Palayamkottai.

Taxonomical Classification

Kingdom	:	Plantae, Plants
Sub Kingdom	:	Tracheobionta, Vascular plants;
Super Division	:	Spermatophyta, Seed plants;
Division	:	Angiosperma
Class	:	Dicotyledonae
Sub class	:	Gamopetalae

Series	:	Bicarpellatae
Order	:	Personales
Tribe	:	Justicieae
Family	:	Acanthaceae
Genus	:	Andrographis
Species	:	A. paniculata(Burm, f) Nees ²⁰

Vernacular names¹¹⁶⁻¹¹⁷

• criticatar fi	unico	
Arabic	:	Quasabhuva
Assamese	:	Chiorta, Kalmegh
Azerbaijani	:	Acılar Şahı, Acılar Xanı (khanı)
Bengali	:	Kalmegh
Burmese	:	Se-ga-gyi
Chinese	:	Chuan Xin Lian
English	:	The Creat, King of Bitters
French	:	Chirette verte, Roi des amers
Gujarati	:	Kariyatu
Hindi	:	Kirayat, Kalpanath,
Indonesian	:	Sambiroto, Sambiloto
Japanese	:	Senshinren
Kannada	:	Nelaberu
Konkani	:	Vhadlem Kiratyem
Lao	:	La-Sa-Bee
Malay	:	Hempedu Bumi, Sambiloto
Malayalam	:	Nelavepu, Kiriyattu
Manipuri	:	Vubati
Marathi	:	Oli-kiryata, Kalpa
Mizo	:	Hnakhapui
Oriya	:	Bhuinimba
Panjabi	:	Chooraita
Persian	:	Nain-e Havandi
Philippines	:	Aluy, Lekha and Sinta
Russian	:	Andrografis
Sanskrit	:	Kalmegha, Bhunimba and Yavatikta
Scandinavian	:	Green Chiratta
Sinhalese	:	Hīn Kohomba or Heen Kohomba
Spanish	:	Andrografis
Tamil	:	Nilavembu
Telugu	:	Nilavembu
Thai	:	Fa-Talai-Jorn, Fah-talai-jon (jone)
Turkish	:	Acılar Kralı, Acı Paşa, Acı Bey
Urdu	:	Kalmegh, Kariyat, Mahatita
Vietnamese	:	Xuyên Tâm Liễn

MORPHOLOGICAL CHARACTERS OF A. PANICULATA118-119

The leaves and roots of *A. paniculata* have been traditionally used in Asia and Europe over the periods for different medicinal purposes as a folktale remedy for a wide spectrum of ailments⁶¹. The plant generally grows up to a height of 30-110 cm in moist shady places. The morphological appearances of leaves are simple, dark in color, opposite, lanceolate, glaborous, 2- 12cm long, 1- 3cm wide, acute apex, and entire margin²². The stem is acutely quadrangular or Tetrangular in outline; can be broken easily due to its fragile texture and it is much

branched. Roots showed the presence of secondary growth which is visible. Inflorescence of the plant is 10-30 mm long; bract small; pedicel short.^{20,24-27} Flower consists of small, linear 5 particle, calyx; tube narrows, about 6mm long white corolla with violet markings. Two stamens inserted in the throat and two celled superior ovary.1-2cm long, 2-5mm wide, linear-oblong, compressed, erected capsule²⁸. Seeds are numerous, sub quadrate and yellowish brown. Micropropagation is the proven method for efficient *in vitro* propagation of medicinal and aromatic plants and for commercial exploitation of valuable plant-derived pharmaceuticals²⁹⁻³⁵.

MORPHOLOGY OF ANDROGRAPHIS PANICULATA



Figure 1 A) Mature seeds B) Leaves C) Aerial parts with mature and immature capsule; D) Flower buds E) Entire flower F) Mature Fruit (Photo courtesy: Ankita kataky, 2010).

Table 1
Morphological characters of Andrographis paniculata.

1	Plant height	30–110 cm
2	Stem	Dark green
	Length	30–100 cm
	Diameter	2–6 mm
	Shape	Quadrangular with longitudinal furrows and wings on the angles of the young parts, slightly enlarged at the nodes
3	Leaves	Glabrous
	Length	2–12 cm
	Width	1–3 cm
	Arrangement	Lanceolate
	Shape	Pinnate, acute apex, entire margin
4	Flowers	White with rose-purple spots on the petals
	Size	Small, in lax spreading axillary and terminal racemes or panicles
5	Seed	Capsules linear-oblong, acute at both ends
	Size	1.9 cm × 0.3 cm
	Color	Yellowish brown
	Shape	Subquadrate, numerous
6	Flowering and fruiting	December to April

MICROSCOPICAL STRUCTURE OF A. PANICULATA

Angles of A. paniculata stem are short with winged projections which consists of single layer epidermial cell and a group of parenchymatous cells; having the ridges in between which are shallow; compactly arranged cells with a thin cuticle and it is allied with glandular hairs externally; 2-5 cell layer thick walled hypodermis, cells more or less rounded or polygonal, thin, compactly arranged with the presence of chlorophylls; cortical cells are 5 to 6 layer thick, rounded, thin walled, compact, and parenchymatous; stele amphipholic siphonostele subjugating supreme part of the stem, spreaded more to the ridge area a few sclerenchymatous cell in the periphery of vascular bundles groups comprises of 2 to 4 or even solitary throughout; external phloem is thin layered whereas internal phloem occurs mostly in patches; xylem disturbed with medullary rays; pith parenchymatous, cells polygonal to rounded, larger in size, thick walled, and compactly arranged without any

content²³.Roots of epiblema is replaced by cork layer, three different separate zones of which cells of peripheral layer are arranged loosely, inner cells are smaller but arranged compactly; stele occupies the maximum part of the root being the secondary growth; xylem and ray cells are distinct in rows, primary vascular crushed: pith insignificant having bundle few parenchymatous smaller cells²³. The leaves of A. Paniculata showed the presence of diacytic stomata at lower epidermis, trichomes may be glandular or nonglandular, large cystoliths, columnar palisade cells, beneath epidermal layer, collenchymas are present in midrib; spongy parenchyma cells; vascular bundles are spiral, scalariform and reticulate lignified xylem vessels in the upper part and lignified phloem in the lower part with small acicular calcium oxalate crystals, a film of lower epidermal cells showed wavy-wall, dense collenchyma at the corners of stems, a layer of thickincorporates chloroplastid.²⁸ parenchyma cells

MICROSCOPICAL SECTION OF A. PANICULATA



Figure 2

Matured erect branched plant; 2- Flower lobes showing pigmentation; 3- Dark purple anther;
4- Fruits; 5- Golden brown seeds; 6- T.S. of stem; 7- T.S. of root; 8-Stained and unstained pollen grains; 9- Metaphase I (Datta Kumar, 2012).

NURSERY

For the cultivation of A. paniculata in a small scale nursery, vermicompost coir pith are used for the recovery of soils from industrial sites³⁶. Seeds were collected and sown in polythene bags. The nursery beds were prepared for three treatments with biocompost. vermicompost, farmvard manure and one bed for control. The treatments were added to soil in 1:2 ratios. After few days of seed germination plantlets were transplanted in respective beds. After every two months three samples were taken from each bed. The plantlets were uprooted. Increased shoot length was observed in vermicompost treatment (10.11cm) followed by farmyard manure, biocompost and control. The increase in growth may have been due to increases in microbial biomass in soils receiving vermicomposts which increased nutrient mineralization

SOIL CONDITIONS

It is cultivated in soil types, particularly serpentine soils which are legitimately high in metals such as copper, zinc and aluminium and no other plant can be cultivated in such soils³⁷. Wet soil or flooded soil is side stepped for its cultivation³⁸.

HARVESTING PERIODS

A. paniculata is upraised from seeds and favors sunny condition. The seedlings elevated in nursery beds should be transplanted to field at a distance of 60 cm \times 30 cm with three irrigations during that period, predominantly at flowering stage. In India, the seeds are sown in the months of May – June, flowers during August – November and the whole plant starts maturing during February – March. Maximum harvest of total diterpene lactones was noted as flourishing from the aerial part³⁹⁻⁴¹. The best harvesting period of *A. paniculata* leaves is at 3-5 months old or at 50%

blossom where the highest quantity of active lactone compounds were present followed by final harvesting after next 2-3 months, with an yield of 3 ton per hectare (fresh weight) or 0.5-1 tons per hectare (dried weight)⁴².

PROPAGATION

After 6 weeks of sowing, the seedlings were transplanted in the main field at the spacing of 15 x 15 cm. The crop were uniformly applied with 75 kg each of Nitrogen, Phosphorous and 50 kg of Potassium per hectare and cultural operations like irrigation, weeding were practiced. The propagation of *A. paniculata* generally occurs through the seeds. In the natural habitat, it is found growing in clay to sandy loamy soil rich in organic matter is good for its growth and yield¹³⁷. After the application of FYM positively influenced seedling growth of kalmegh in the nursery. At transplanting age (47 DAS) highest seedling height with maximum number of leaves was obtained with 14 kg/m² FYM application⁴⁸.

GENETIC MALE STERILITY

Genetic male sterility induced (6.0 to 14.0%) in *A.* paniculata at M_1 following 20 kR gamma irradiation was reported by Lattoo *et al.*,(2006) ⁴³monogenic recessive to normal was found in the male sterile gene. It acted upon the tapetal layer and also affects the non sporogenous tissue within the anther locule resulting in infringement of the locule and thereby, significantly it reduces the production of pollens and enhances the creation of abortive pollen. However, female fertility in *A. paniculata* remained unimpaired and completely intact.

TISSUE CULTURE TECHNIQUES

The regeneration of shoot buds from organogenic calli was varied on the basis of the culture medium composition. About 75.3% in case of leaf-derived calli and 63.4% in case of stem-derived cultures showed shoot bud regeneration in the medium having 3.0 mg/L BA, 50 mg/L Ads and 1.0 mg/L NAA after six weeks of first subculture. The increase of NAA concentration higher than 2.0 mg/L suppressed the rate of shoot bud regeneration and slow growth of the organogenic calli. The maximum number of shoot buds (28.6) was obtained in the medium containing 3.0 mg/L BA, 50 mg/L Ads and 1.0 mg/L NAA after four weeks of culture initiation. A high frequency of shoot production from organogenic calli could be obtained by manipulating the growth regulators and culture condition. There were differences between treatments both in the percentage of cultures with response and in the mean number of shoot buds per culture.Effect of BAP (1.0 µM) in combination with other cytokinins (0.5-10.0 µM Kn, 2iP and TDZ) were tested and results showed synergetic effect of cytokinins and induced number axillary shoots. The results were best at BAP (1.0 µM) in combination with Kn (5.0 µM) and 39.08 shoots developed after six weeks of culture. However, combination of BAP + 2iP and BAP + TDZ was not much beneficial in triggering the percentage of responding explants and provoking multiple shoot formation indicating that multiple shoot formation depends on the optimum balance of growth regulator combination and concentration in the medium. Similar differential response was observed¹³⁹

TISSUE CULTURE OF A. PANICULATA.

INDUCTION OF ROOTING AND ACCLIMATIZATION

Elongated shoots were excised and placed in half or full strength MS medium supplemented with various concentrations of IBA or NAA for root induction. Full strength MS medium without growth regulators did not promote root induction; roots were observed in media containing 1/2 strength MS medium supplemented with NAA or IBA with 2% sucrose. However, optimal rooting and growth of micro shoots were observed in medium containing 0.5 mg/L IBA or NAA with 2% sucrose after 9 to 11 days of culture without intervening callus. The maximum percentage of rooting (76.2%) was obtained in the medium containing 0.5 mg/L IBA within three weeks of culture. Root development was however, slow at higher concentrations of IBA or NAA. The rooted plantlets were transferred into pots for acclimatization. About 60% of the rooted plantlets survived in the pot one week after the transfer and then the plants were grown normally. A successful production of shoot bud regeneration from leaf and stem explants and induction of roots from excised root were dependent on the nutrient medium and the culture conditions.



Figure 3

A) Organogenic calli development from leaf explant on medium having 3.0 mg/L BAP + 50 mg/L Ads + 1.0 mg/L NAA after 6 weeks of initial culture. B) Shoot bud regeneration (arrows) from Organogenic calli on medium having 3.0 mg/L BAP + 50 mg/L Ads + 1.0 mg/L NAA after 6 weeks of subculture. C) Root initiation from micro shoots of Andrographis paniculata after 3 weeks of culture of ½ strength MS medium supplemented with 0.5 mg/L IBA + 2% sucrose. D) Plantlets established in the pot. (Picture courtesy - Bansi and Rout (2013)).Compounds in A. paniculata

The plant extract is known to contain diterpenes, flavonoids and stigmasterols. Flavonoids are present more in roots and leaves. From the soluble fractions of the ethanol or methanol extract, 5-hydroxy-7,8dimethoxyflavone, 5-hydroxy-7,8,2',5'tetramethoxyflavone, 5-hydroxy-7,8,2',3'tetramethoxyflavone, 5-hydroxy-7,8,2',3'tetramethoxyflavone, 7-O-methylwogonin and 2'-methyl ether were isolated as the main flavonoids. "The aerial parts contain alkanes, ketones, and aldehydes.

Andrographolide is the key constituent having a molecular formula as $C_{20}H_{30}O_5$, and is believed to be the active constituent for biological activities and represents as an identity indicator for the plant. Melting point of Andrographolide is 230°-239°C. (3 - (2 - decahydro -6 - hydroxyl - 5 - (hydroxymethyl) - 5, 8a - dimethyl -2-methylenenaphthyl) ethylidene) dihvdro-4hydroxyfuran- 2 (3H) - one) is the reported IUPAC name of andrographolide. The compound Andrographolide can be easily dissolved in methanol,

ethanol, pyridine, acetic acid and acetone, but slightly dissolved in ether and water. Growing region and seasonal changes have a strong impact on formation of

the diterpene lactones. The highest concentration of the active components is found just before the plant blooms, making early fall the best time to $harvest^{44\&20}$.

Molecular structure of Andrographolide



The other important compounds isolated from different parts of A. paniculata are apigenin-7, 40-di-omethyl ether, carvacrol, eugenol, myristic acid, hentriacontane, tritriacontane, or oxylon A and wogonin⁴⁵. the chemical composition of Andrographis paniculata showed that it is a rich source of deterpenoids and 2'-oxygenated flavonoids including andrographolide. neoandrographolide, 14-deoxy-11,12didehydroandrographolide, 14-deoxyandrographolide, isoandrographolide and 14-deoxyandrographolide 19 β-D-glucoside, homoandrograholide, andrographosterin, stigmasterol^{46&48;21} andrographan, andrographiside, deoxyandrographiside, homoandrographolide, andrographan, andrographon, andrographosterin²⁰.

BIOACTIVITIES OF A. PANICULATA

The plant is used as bitter stimulant, antispasmodic, antiperistaltic, stomachic and also an antihelmintic. It has been active with various advantages in case of general weakness in recuperation after fevers, disorders of liver and advanced stages of dysentery⁴⁹. The plant is also cast off to treat gastro-intestinal tract and upper respiratory infections, fever, herpes, sore throat, and a variety of other chronic and infectious diseases. The fresh leaves of *A. paniculata* juice of are a domestic remedy in the treatment of colic pain, loss of appetite, irregular stools and diarrhea

BIOACTIVITIES C ANDROGRAPHOLIDE

OF

The bioactivities of some of the andrographolide compounds are as follows;14-deoxyandrographolide responsible for activation of NOS and guanylate cyclase vasorelaxation in vitro and in vivo neoandrographolide - NO, PGE2, iNOS and COX-2 in activated macrophages CCl4, tBHP-induced hepatotoxicity (*i.p* 100 mg/kg, 3d)⁵⁴⁻⁵⁶; 14-deoxy-11,12 didehydroandrographolide - muscle relexation and Nitric oxide release from endothelial cells⁵⁷⁻⁵⁸; 14-deoxy-14,15-didehydroandrographolide - cytotoxic activity and cell cycle arrest of tumor cells NF-kB-dependent transactivation⁵⁹⁻⁶⁰; andrograpanin - protein kinase or p38 MAPKs pathways chemokine SDF-1α induced cells⁶¹⁻⁶²: in Jurkat and THP-1 chemotaxis isoandrographolide - cell-differentiation-inducing activity

proliferation of HL-60 cells $^{63-64}$; 14acetylandrographolide - growth of leukemia, ovarian, renal cancer cells 65 19-O-acetyl anhydroandrographolide - NF- κ B-dependent transactivation 60 .

PHARMACOLOGICAL EFFECTS

Some of the pharmacological effects of andrographis are followed as, Abortifacient which can abort pregnancy. Acrid which is rubifacient to the skin; Analgesic; Antibacterial; Antidiarrhoeal and intestinal effects; Anti-inflammatory activity; Antimalarial activity; Antioxidant activity; Antipyretic; Anti snake venom; Antithrombotic; Antiviral; Cancerolytic; Cardiovascular activity: Choleretic: Depurative: Expectorant: Hypoglycemic activity; Hepatoprotective activity; Immunological potential; Laxative; Psychopharmacological activity; Sedative: Thrombolytic; Vermicidal.²⁰⁻²¹

ANTIOXIDANT EFFECTS

Verma and Vinayak (2008)¹⁴³ related the antioxidant effects of the aqueous extract on liver defense systems in lymphoma bearing mice. The aqueous extract and hydro alcoholic extract of the medicinal plant A. paniculata showed the increase in activities such as catalase, superoxide dismutase and glutathione-Stransferase enzymes and reduced lactate dehydrogenase activity. The results performed with that of aqueous extract of A. paniculata exhibited a greater antioxidant activity than the ethanol extract in all model systems tested. The function of Hydroalcoholic extract of A. paniculata possesses oxidative alterations in myocardium and confers substantial cardioprotective activity by facilitating in retaining the cardiac function in a norma manner⁶⁶.

ANTI-DIABETIC ACTIVITY

Radhika and her coworkers (2012)¹⁴⁴ using carrageenan induced rat hind paw oedema modelfor acute inflammation. The anti-inflammatory activity of chloroform extract of *Andrographis paniculata* stem was determined by Ibuprofen as a standard drug. In 6th hour

at a dose of 200mg/kg the chloroform extract of Andrographis paniculata stem showed good effect and was comparable with the standard drug Ibuprofen (10 mg/kg) $(t = 64.06, p < 0.001)^{67}$. The compound Andrographolide reduces the plasma glucose concentration in streptozotocin induced in both the diabetic rats and in normal rats, having a significant effect in normal rats than on diabetic rats and in addition to it an intravenous glucose is retort to the normal rats and enhances the uptake of radioactive glucose by isolated soleus muscle of streptozotocin-diabetic rats in concentration-dependent manner. Intravenous а administration of andrographolide in diabetic rats for continuous three days indicates that the glucose lowering effect of andrographolide is due to better utilization of glucose by skeletal muscle which results in increase in mRNA and protein levels of glucose transporter (GLUT4) in the soleus muscle.⁶⁸However, after in vitro experiments, Wibudi and his coworkers¹ (2008) determined that the hypoglycemic effect of Andrographis paniculata is due to insulin released from pancreatic *β*-cells through ATP-sensitive potassium channels, similar to other insulinotropic antidiabetic agents⁶⁹.

ANTIDIARROHEAL ACTIVITY

The inflammation can be caused by pathogenic bacteria growth or a viral or parasitic infections and irritations. Medications and certain foods are the sources of pathogenic growth. Campylobacter, Salmonella. Shigella and Escherichia coli are common bacteria that cause diarrhoea. Although antibiotics are effective in treating bacterial infections, antibiotic-resistant strains of bacteria can be produced by the overuse of antibiotics. Many drugs, such as kaolin-pectin, loperamide and bismuth are used to relieve the symptoms of diarrhoea, but they may cause undesirable side effects⁷⁰. The A. paniculata components, Andrographolide and neoandrographolide showed comparable activity to loperamide (Imodium), the most common anti-diarrhoea drug. In Thailand, a methanolic extract was made to boil A. paniculata stem was testified to be effective against Proteus vulgaris and combined powder of stem and leaves can be effective against the Shigella bacteria ⁷¹. In an experiment conducted in a pharmacological research institute in Shanghai, China, 165 patients were given tablets of A.paniculata which is equal to the amount of 15.6g crude powder per day. Fluroxone, a common drug used to treat dysentery were given to 28 patients. The result significantly showed the effective A. paniculata was 75.2% than that of rate of Fluroxone was 71.4%⁷². A. paniculata was believed to be effective against bacterial dysentery and diarrhea because it has antibacterial activities. In a study conducted on mice, it was found that 50% and 85% alcohol extracts of Andrographis paniculata leaf powder were effective in reducing intestinal tract movements⁷³.

ANTIMICROBIAL ACTIVITY

The plant *Andrographis paniculata*, is an antibacterial agent capable of responding the ill effects caused by pathogenic microbes⁷⁴⁻⁷⁷. The antimicrobial activity of

aqueous leaf extract of A. paniculata was found to have antibacterial activity against Bacillus subtilis and Streptococcus aureus by Manjusha et al (2011)⁷⁹. A similar conclusion was reported by Radha et al (2011)⁹⁴ who found that petroleum ether, acetone, chloroform and methanol extracts of A. paniculata leaves and stems, showed significant antimicrobial potential against Enterococcus faecalis, Streptococcus pyogenes, Klebsiella pneumonia and Proteus vulgaris. The ethanolic leaf extract and andrographolide compound isolated from the leaves are potent in inhibiting these bacteria and the work highlights that the inhibitory effect with standard antibiotics. In vitro screening of the aqueous extract of A. paniculata posses' potential antibacterial activity towards both gram-positive and gram-negative microorganisms where the methanolic extracts inhibited the growth of 95% organisms tested, followed by chloroform extracts inhibited 80%. Hexane extracts inhibited 65% growth of the tested organisms⁸⁰ The antimicrobial activity of aqueous extract of Andrographis paniculata or andrographolides and arabinogalactan proteins from A. paniculata when evaluated, showed significant antimicrobial activity, which may be due to the combined effect of the isolated arabinogalactan proteins and andrographolides⁸¹.

ANTICANCER ACTIVITY

Chen and his coworkers (1982)⁴⁸ reported that a major mediator of the inflammatory response is elevated interleukin-6 (IL-6), has been concerned in androgen receptor (AR) activation, cellular growth and differentiation, which plays an important role in the development and progression of prostate cancer, and is a potential target in cancer therapy. Through screening of drug using human prostate cancer cell lines expressing that IL-6 autocrine loop, showed the presence of andrographolide, a diterpenoid lactone isolated from a traditional medicinal plant. Andrographis paniculata, could significantly inhibit and suppresses the IL-6 expression and its mediated signals. Andrographolide inhibits IL-6 expression at both mRNA and protein levels in a dose-dependent manner and it suppresses both IL-6 autocrine loop and paracrine loop induced cell signaling including Stat 3 and Erk phosphorylation. Furthermore, andrographolide hinders cell viability and induces apotopsis in both androgen stimulated and castration resistant human prostate cancer cells deprived of causing substantial toxicity to normal immortalized prostate epithelial cells. Moreover, treatment of andrographolide to mice bearing castrationresistant DU145 human prostate tumors that express constitutive IL-6 autocrine loop significantly suppresses growth. These results demonstrate tumor that andrographolide could be developed as a therapeutic agent to treat both androgen stimulated and castration resistant prostate cancer possibly by suppressing IL-6 expression and IL-6 induced signaling⁸².Andrographolide is the key compound used in the treatment which inhibited the in vitro proliferation of different tumor cell lines, signifying numerous types of cancers. The compound exerts direct anticancer activity by cell-cycle arrest at G0/G1 phase through induction of cell-cycle inhibitory protein p27 and decreased expression of cyclin-dependent kinase 4 (CDK4).

Immunostimulatory activity of andrographolide is demonstrated by increased proliferation of lymphocytes and production of interleukin-2. Andrographolide also enhanced the tumor necrosis factor- α production and CD marker expression, resulting in increased cytotoxic activity of lymphocytes against cancer cells, which may contribute for its indirect anticancer activity. The *In vivo* anticancer activity of the compound is further substantiated against B16F0 melanoma syngenic and HT-29 xenograft models and the results suggest that andrographolide is an interesting pharmacophore with anticancer and immunomodulatory activities and hence has the potential for being developed as a cancer therapeutic agent.

ANTI-PLASMODIAL ACTIVITY

In vitro studies of Dua and his coworkers (2004)¹⁴⁶ revealed that compound 1,2-dihydroxy- 6,8-dimethoxyxanthone possessed substantial anti-plasmodial activity against *Plasmodium falciparum* with its IC_{50} value of 4 µg ml⁻¹. Xanthones bearing hydroxyl group at 2 positions demonstrated most potent activity while xanthones with hydroxyl group at 1, 4 or 8 position possessed very low activity. *In vivo* antimalarial sensitivity test of this compound on Swiss Albino mice with *Plasmodium berghei* infection using Peters' 4-day test gave substantial reduction (62%) in parasitaemia after treating the mice with 30 mg kg-1 dose⁸⁴ The methanolic extract significantly inhibited *Plasmodium falciparum* at a 50-percent inhibitory concentration (IC50) of 7.2µg/mL⁸⁵.

ANTICOLD ACTIVITY

One clinical trial has investigated the efficacy of a standardized *A. paniculata* extract to prevent the common cold by Caceres 107 healthy students in a rural school had daily taken either placebo or a dose of 200 mg (minimum 5.8%) of Kan Jang (a formulation of *A. paniculata* provided by the Swedish Herbal Institute) for three months. The number of colds occurring over a three month period was observed. After 1 month no significant difference was found. However, the difference was statistically significant in the second and third month. The placebo group was 2.1 times more likely to catcha cold than the Kan Jang group. The incidence of the common cold was 30% in the *A. paniculata* group, whereas the incidence was 62% in the placebo group⁸⁶.

DYSENTERY OR GASTROENTERITIS

Akbar $(2011)^{110}$ reported that the Ethanol extract tablets of *A. paniculata* were reported to cure 88.3 percent of acute bacillary dysentery and 91.3 percent of acute gastroenteritis cases. Andrographolide administration was reported to cure 91 percent of acute bacillary dysentery cases.¹⁴⁰

ANTI-INFLAMMATORY EFFECT

Deng et al (1982)¹⁴⁷ also suggested A. paniculata might exert through stimulation of the adrenal gland since the herb showed no effect when the adrenal gland of the animals were totally removed⁸⁷. Hidalgo et al (2005)⁸⁹ suggested its anti-inflammatory effect involved Andrographolide inhibiting a nuclear factor kappa B (NFkappaB) binding to DNA in endothelial cells or HL-60derived neutrophilic cells, and thus decreasing the expression of proinflammatory proteins ^{88&89}. In a study from Thailand, rats were given injections with carrageenan (an agent for stimulating inflammation) to study the anti-inflammatory effect of A. paniculata extract ranging from 500-1250, 2500 mg/body weight. The result showed that aqueous extract of A.paniculata effectively reduced the paw volume in rats treated with A.paniculata whereas the control group did not Amroyan et al (1999)⁹¹found that Andrographolide from Andrographis paniculata did not affect the biosythesis of eicosanoids, but inhibited the plateletactivating factor (PAF) induced human blood platelet aggregation where eicosanoids and PAF are two of the most important inflammatory mediators. Inhibition of the biosythesis of eicosanoids is characteristic for non steroidal anti-inflammatory drugs, while PAF antagonists are used as potential agents in inflammation. This mechanism is most likely combined with the cardiovascular and antithrombic activity of A. paniculata

ANTIVIRAL ACTIVITY

Andrographolide, neoandrographolide and 14-deoxy-11,12- didehydroandrographolide are reported to be viricidal against herpes simplex virus 1 (HSV-1) without having any significant cytotoxicity at viricidal concentrations⁹³.

ANTI- HIV ACTIVITY

Stephen and Comac (2000)¹⁴⁸ indicated that extracts of *Andrographis paniculata* may have the potential for interfering with the viability of the Human Immuno Deficiency Virus (HIV) and advised that *A. paniculata* could combine with modern medicines against Acquired Immuno Deficiency Syndrome (AIDS) A phase I dose-escalating clinical trial conducted in 13 HIV patients showed a significant rise in the mean CD4(+) lymphocyte level but with no significant changes in mean plasma HIV-1 RNA levels of HIV- 1 infected patients after administration of the regimen. The findings proved that andrographolide may inhibit HIV-induced cell cycle dysregulation leading to a rise in CD4 (+) lymphocyte levels in HIV-1 infected individuals⁹².

ANTI-FUNGAL ACTIVITY

Radha *et al* (2011)⁹⁴ examined the petroleum ether, acetone, chloroform and methanolic extracts of *Andrographis paniculata* leaves and stems, in order to evaluate the antifungal potential of *Candida albicans* and *Aspergillus flavus*. The yeast, *Candida albicans*

showed susceptibility to 75% of chloroform extracts of the leaves (23.33; 1.20mm) and the acetone extracts of stems showed inhibitory effect on the growth of the fungus, *Aspergillus flavus* (23.67; 0.88mm)^{79.} Similar studies were conducted by Bobbarala *et al* (2009)⁸⁰. against *Acremonium strictum*, *Alternaria alternata*, *Aspergillus flavus*, *Bipolaris bicolor*, *Cladosporium herbarum*, *Curvularia lunata*, *Fusarium oxysporum*, *Penicillium expansum*, *Rhizoctonia solani*, *Tiarosporella phaseolina* and *Ustilago maydis* using hexane, chloroform and methanolic extracts and the results revealed that the methanolic extracts showed activity against *Alternaria alternate* whereas, the chloroform extracts showed greater activity against *Fusarium oxysporum*⁹⁵.

OTHER ACTIVITIES

Xanthones isolated from the roots showed antiprotozoal activity against Trypanosoma brucei, Trypanosoma cruzi and Leishmania infantum⁹⁶. The compound 1,2dihydroxy-6,8-dimethoxy-xanthone possessed substantial antiplasmodial activity against Plasmodium falciparum with its IC₅₀ value of 4 µgml⁻¹. Xanthones bearing hydroxyl group at 2 position demonstrated most potent activity while xanthones with hydroxyl group at position possessed 1.4 or 8 verv low activity⁸⁴. Nematicidal efficiency of water and methanolic extracts of Andrographis paniculata were evaluated In vitro against root knot nematode, Meloidogyne javanica and reniform nematode, Rotylenchulus reniformis by Goel et al (2009).96 Crude fractions of aerial parts of Andrographis paniculata were evaluated for growthinhibitory and oviposition-deterrent activity against larval and adult stages of Bihar hairy caterpillar, Spilarctia oblique (Arctiidae). The methanol fraction had the highest growth inhibitory activity. The diterpene andrographolide, displayed significant growth-inhibitory antifeedant properties with GI50 and FD50 values of 100.4 and 159.7 µg/g diet respectively. The ethyl acetate fraction possessed the highest ovipositiondeterrent activity⁹⁸. Andrographis paniculata including andrographolide, a major diterpenoid component and its analogues have been reported to exhibit a marked effect on hepatic bio-transformation enzymes, i.e., aniline hydroxylase, N- and O-demethylase⁹⁹alanine aminotransferase and aspartate aminotransferase¹⁰⁰including phase II enzymes, i.e., glutathione S transferase and DT-diaphorase¹⁰¹. Modulatory influence of Andrographis paniculata extract on a responsive isoform of hepatic CYPs was recently reported in mouse hepatic microsomes compared to typical CYP-inducers (3-MC for CYP1A and PB for CYP2B), in terms of total CYP content and related alkoxyresorufin O-dealkylase activities¹⁰²

HYPOTENSIVE ACTIVITY

Andrographis paniculata, is reported to have, by acting through β - adrenoceptors, autonomic ganglion receptor and angiotensin converting enzyme (ACE) inhibitory activity¹⁰³. The 4th week of the extract treatment in SH rats significantly increases the relaxation responses to ACh as a result of possible improvement in the

endothelial function, these are comparable with the study. Conversely the plant possesses a remarkable capability to challenge the nor-epinephrine induced contractions resulting in vasorelaxation in isolated rat¹⁰⁴. The improvement in relaxation responses to ACh following chronic administration of chloroform extract is most likely due to the activation of NO synthase and ultimate stimulation of NO production in endothelial cells. Moreover, the effects of chronic administration are evidently suggestive of increased responsiveness of the vascular smooth muscle to NO since 4-week treatment with the extract was found to enhance the relaxation responses to the action of the endothelium-independent vasodilator SNP. Endothelial protective effects of Andrographis paniculata chloroform extract were comparable to the effects of Verapamil, which acts by blocking the L-type Ca2+ current and high K+ activated pathways to relax the smooth muscle¹⁰⁵

ANTIVENOM ACTIVITY

Plant extracts (7.2 mg/kgbody weight) and partially purified fractions (2.4mg/kgbody weight) when orally administered to mice experimentally envenomed with rattle snake venom injection (2.5- 15 mg/kg body weight) showed potent neutralizing effect against the venom. The isolated fractions effectively inhibited the toxic effect of snake venoms *in vitro* than *in vivo*.¹³⁸

BIOFILMS

The antibiofilm activity of the extract on the exopolysaccharide (EPS) was determined using a modified gradient plate technique with Congo red medium P. aeruginosa isolates were grown in plant extract incorporated trypticase soy broth for 18 hours and allowed to form a biofilm. Water and tobramycin were the negative and positive controls, respectively. After incubation at 37°C, they were stained with crystal violet (0.1%) and quantified using microtitre analysis (OD 490 nm). A continuous gradient of plant extract on CRA medium was prepared, and the biofilm forming isolates were inoculated into the centre of the plate as a single streak. The organisms were again streaked zigzag perpendicular to the original streak crossing it each time. The colour and nature of the developed colonies along the streaked line were observed¹⁰⁶. The crystal structure of P. aeruginosa 3JPU (P. aeruginosa LasR, transcriptional regulator of Las QS system), 2B4Q (RhIG, beta-ketoacyl reductase, QS regulated virulence factor rhamnolipid synthesis enzyme) and 2W38 (P. aeruginosa sialidase, in vivo biofilm formation enzyme) were retrieved from the NCBI-PubChem database. Protein-compound docking simulation was performed using AutoDock 4.0¹⁰⁸. The ADME molecular properties and bioactivity scores of the drug targets were calculated using Molinspiration according to Lipinski's rule for all analysed ligands¹⁰⁹.

MACROMUTANT ACTIVITY

Ghosh *et al* (2012).²³ screened 14 viable (*S. viridis, lax branching, bushy, unbranched I* and *II, dark green leaf, broad leaf I* and *II, narrow leaf I* and *II, drooping leaf I*

and *II*, *dwarf* and *early maturity*) at M₂ following EMS and dES treatments. Mutation frequency over M₂ population was 2.82% and *lax branching* mutant was maximum (0.51%). EMS induced relatively higher (3.12%) frequency of mutation than dES (2.46%). The mutant traits were monogenic recessive mostly (*S. viridis* showed digenic mode of inheritance). All mutants bred true in M₄ generation. Meiotic analysis revealed 2n=50 chromosome in mutants alike to control. Andrographolide (estimated from matured leaves by HPTLC) content (control: 3.41%, mutants: 0.03 to 3.99%) was significantly higher in *bushy* and *broad leaf I* and *II* than normal plants. The mutants induced were considered to be important genetic resources in the plant species.

MOLECULAR WORKS OF ANDROGRAPHIS PANICULATA

In plants, plastidial 2C-methyl-D-erythritol-4-phosphate (MEP) and cytosolic mevalonic acid (MEV) pathways provide two 5C isoprenoid building blocks, dimethyl allyl

Proposed pathway for ent-LRD biosynthesis in kalmegh.

diphosphate (DMAPP) and isopentenyl diphosphate (IPP), for the biosynthesis of diverse terpene metabolites¹²⁰. IPP and DMAPP derived from the MEP pathway are converted to monoterpenes, diterpenes, and tetraterpenes, whereas those derived from the MEV pathway are converted to sesquiterpenes and triterpenes. However, cross-talk between these two pathways in biosynthesis of some terpenes was also recognised ¹²¹⁻¹²³. Previously, a major role of the MEP pathway and a minor role of the MEV pathway for supplying the 5C isoprenoid precursors for the biosynthesis of andrographolide were reported¹²⁴. Transcripts predicted to encode all the enzymes of the MEP and MEV pathways are identified in kalmegh transcriptome which demonstrates the quality and indepth coverage of the transcriptome database. Interestingly, four transcripts for DXS, three for DXR and two each for HDS, HDR, AACT, HMGS, HMGR, PMK, MVD and IDI were revealed in kalmegh transcriptome. This observation suggests the likely existence of multiple isomers for these enzymes in kalmegh.



Figure 4

Putative transcripts of the pathway and corresponding enzymatic steps are shown. AD, andrographolide; NAD, neoandrographolide; DDAD, 14-deoxy-11, 12-didehydroandrographolide (Photo courtesy –Anchal Garg et al., 2015).

This article can be downloaded from www.ijpbs.net B - 193 The second stage of diterpene biosynthesis involves head-to-tail condensation of three IPP and one DMAPP to a C20 compound geranylgeranyl diphosphate (GGPP). Thus, based on structures of kalmegh ent-LRDs, the involvement of class I and class II diTPSs, CYP450s and GTs enzymes in the biosynthesis of ent-LRDs was hypothesized. From kalmegh transcriptome database, three partial transcripts for the GGPS and three full-length transcripts for class II diTPSs with homology to the ent-copalyl diphosphate synthase (ent-CPS) are identified. Besides, several transcripts for CYP450s and GTs are also recognized. Transcripts that encode MEP pathway enzymes, GGPS, class II diTPS, CYP450 and GT, and preferentially expressed in leaf tissue are potential candidates for the biosynthesis of ent-LRDs in kalmegh. Although, two class I diTPSs with sequence similarity with ent-kaurene synthase are identified none of them preferentially expressed in leaf. Thus, their involvement in the biosynthesis of ent-LRD medicinal compounds in kalmegh may be excluded.

IDENTIFICATION OF SIMPLE SEQUENCE REPEATS IN DITERPENE BIOSYNTHETIC PATHWAY TRANSCRIPTS

Simple sequence repeats (SSRs) are often considered most efficient and reliable molecular markers for detecting genetic variations in plants¹²⁵. A total of 16,485 potential SSRs were identified in 13,805 leaf transcripts whereas, 15,911 SSRs were detected in 13,213 root transcripts. Moreover, 2194 leaf and 2200 root transcripts were detected with more than one SSRs. Di-nucleotide repeats were the most abundant SSRs in leaf and root transcripts with 5194 and 5023 SSRs, respectively. The numbers of compound SSRs were 1877 and 1895 in leaf and root transcripts, respectively. Interestingly, several SSRs were also identified in transcripts of the specialized metabolic pathways, including terpenes and phenylpropanoids. SSRs were detected for the transcripts of the MEP pathway enzymes (DXS, MDS and HDR), GGPS and class II diTPSs (ApCPS2, ApCPS3). These SSRs could be useful in genotyping cultivars and developing specific chemotypes of kalmegh following marker-assisted selection.

IDENTIFICATION AND ANALYSIS OF DITERPENE SYNTHASES

Annotation of the kalmegh transcriptome revealed three diTPSs that showed close phylogenetic relationship with the dicotyledons monofunctional class II diTPSs of ent-CPP product specificity. These are ApCPS1 (ApU55291), ApCPS2 (ApU48901) and ApCPS3 (ApU53774). Similar to class II diTPSs, the highly conserved DXDD motif that is essential for the protonation-initiated cyclization of GGPP was identified in ApCPS1, ApCPS2 and ApCPS3, following multiple sequence alignment. Sequence analysis revealed that ApCPS1, ApCPS2 and ApCPS3 encode for 832-, 817- and 797- amino acids proteins with calculated molecular masses of 95.45, 93.43 and 90.81 kD,

respectively. At the amino acid sequence level, ApCPS1 shared 55.2 and 57.21 % identities with ApCPS2 and ApCPS3, respectively. However, ApCPS2 shared 63.36 % amino acid identity with ApCPS3. Like other plant diTPSs, N-terminal transit peptides for the chloroplast localization recognised were in ApCPS1, ApCPS2 and ApCPS3. But these exhibited dissimilar expression patterns in leaf and root tissues. The transcripts levels of ApCPS1 were comparable in leaf and root tissues. However, ApCPS2 showed high level of transcript accumulation in leaf and low level of transcript accumulation in root. ApCPS3 transcripts are present at very high level in root and at very low level in leaf. This divergent expression pattern of ApCPS1, ApCPS2 and ApCPS3 specied their role in different diterpene metabolic pathways of kalmegh, although, their involvement in same biosynthetic pathway with functional redundancy cannot be completely excluded. In order to determine potential of ApCPS1, ApCPS2 and ApCPS3 in functions kalmegh, transcripts levels were analysed to correlate transcript expression with metabolite accumulation pattern, the level of andrographolide, the most abundant ent-LRD of kalmegh, was determined in plant organs and during seedling developmental ages following HPLC analysis. Maximum transcript level for ApCPS1 was detected in stem (4.03-fold), followed by seedlings at cotyledonary leaf stage (CLS, 2.79-fold) as compared to germinating seeds (GS). ApCPS1 transcript was also detected during seed germination. For the biosynthesis of phytohormone gibberellin (GA) ent-CPP serves as precursor which is known to promote seed germination, seedling development and stem elongation in plant species ^{128, 129, 126, 127,} In contrast to *ApCPS1*, *ApCPS2* transcript expression was maximum in leaf (104.34-fold), followed by stem (14.19-fold) as compared to GS. However, very low level of ApCPS2 transcript was detected during seed germination and in seedlings at the CLS stage, as compared to leaf and stem. Based on the transcript expression and ent-LRD metabolite accumulation patterns in plant organs and during seedling developmental ages, the role of ApCPS2 in tissuespecific accumulation of medicinal ent-LRDs was anticipated. Thus, it is hypothesized this function of ApCPS3 because class II diTPSs are known to play role in root phytoalexin biosynthesis in plants 130 Moreover, kalmegh transcripts putatively encoding momilactone-A synthase, a phytoalexin biosynthetic pathway enzyme¹³³, also expressed at high level in roots.

ELEMENTAL ANALYSIS OF ANDROGRAPHIS PANICULATA

Elemental Analysis of plant samples has shown that the plant sample is a rich source of potassium, calcium, Magnesium, iron, aluminum, sodium and manganese are the chief constituents of the plant. Potassium has 14527 part per million; calcium has 3229 part per million. Very important constituent of the extracellular fluid is Potassium which is the principal intracellular cation, diuretic, ionic balance of the human body and maintains tissue excitability. Calcium (Ca) imparts strength and rigidity to bones and teeth. Calcium ions are also needed in neuromuscular transmission. excitability of nerves, and normal excitability of heart, clotting of blood and promoting muscular contraction. The concentration of sodium is less i.e. 94 ppm (in wild), 96.2ppm (in cultivated). Sodium and potassium help in formation of gastric juice in stomach¹³⁴. Iron (Fe) and zinc is used to make tendons and ligaments and is important for maintaining healthy immune system. Iron is essential for blood as it is an essential part of haemoglobin. Its deficiency can cause anaemia. Aluminum (AI) is now thought to be involved in action of a small number of enzymes. Magnesium (Mg) prevents from some heart disorders and high blood pressure and is associated with improved lung function. It helps in absorbing calcium and phosphorus. It is essential to control insulin levels in blood and is injected in veins in acute heart or asthma attack situations. Magnesium is effective in treating numerous heart/lung diseases. Trace elements such as Manganese, Iron and Zinc are essential in enzymes metabolism. The concentration of zinc is found 67 ppm in both the plant sample. Zinc (Zn) help in growth and repair of body tissues.¹³⁵Among the trace elements the concentration of Molybdenum is found 264 parts per billion. The concentration of selenium varies 313 (wild), to 412 (cultivated) ppb. It showed that the plant holds tremendous promise in providing the variable secondary metabolites and mineral supply that could enhance the curative process of ill health. This plant is suitable to meet the human body requirement as an important supplement¹³⁶. The analysis of different elements in the both the plant sample of Andrographis paniculata indicates that the sample possesses same types of elements but in different concentrations. The variation in concentration can be accounted due to the locality factors viz. soil composition, moisture contents, topography aspects, solubility of minerals diffusion and osmosis traits of the plants. Hence, the difference in concentration of the various elements is attributed to the nature of the plant as well as its locality factors. Furthermore, this difference can also be attributed to the edaphic factors along with the forest management practices. The different quantities of different elements under the present physical conditions of wild and cultivated sites are not in consonance with each other. These inorganic elements play an important role in physiological process involved in human health.

SAFETY

Burgos *et al* (2003)⁵¹ found no subchronic testicular toxicity in male rats treated with the standardized dried extract of *Andrographis paniculata* as evaluated by reproductive organ weight, testicular histology, ultrastructural analysis of leydig cells and testosterome levels after a period of 60 days treatment.In mice that received oral extracts of *Andrographis paniculata* (10kg

body weight) once a day for seven days, could survive and none of the mice died. Heart, kidney, liver and spleen were found to be normal in these animals. When 500mg/kg of *Andrographis paniculata* were given daily for ten days to mice, there was no effect on growth, appetite and stool production. The animals were energetic and results of complete blood counts were normal. As with all herbs, some people will have an allergic reaction to *Andrographis paniculata*.¹⁴¹

CONCLUSION

The plant A. paniculata has been valued for treating various infectious diseases and which are highly showing preventative effects against ailments like liver damage, hyperglycemia, dysentery, cancer, pulmonary tuberculosis, AIDS, acute and common cold, flu, myocardial infarction, inflammation, blood clotting etc. Andrographolide, a diterpenoid lactone having a diversity of pharmacological effects specified in ayurveda, unani, sidhha and traditional chinese medicine system. This herb. It has no toxic effects but yet it found unsafe during the pregnancy. In addition to it a great number of pharmaceutical uses, of which andrographolide has some side effects like nausea, vomiting, loss of appetite which can only be seen upon overdosing. Therefore, researchers may further be undertaken to develop potent formulations consisting of A. paniculata and its isolated molecule, andrographolide by making use of novel herbal drug delivery systems like microparticles. vesicular svstems or through complexation with lipid or other suitable novel carrier.

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CONFLICT OF INTEREST

The four authors (S. Elumalai, R. Banupriya, T. Sangeetha and S. Madhumathi) has declared that there is no conflict of interest with reference to this article.

CONTRIBUTION OF AUTHOR

The Corresponding author Dr. S. Elumalai was the brainchild who motivated to review the article on the medicinally important plant *A. paniculata* and made necessary corrections with the same. The Second and third author R.Banupriya and T.Sangeetha collected the necessary informations and wrote the review article. Followed by them, S.Madhumathi rendered the technical support.

REFERENCES

- 1. Bowers MD, Puttick GM. Fate of ingested iridoid glycosides in lepidopteran herbivores. J Chem Ecol.1986;12:169-178.
- Atal CK, Kapur BM. Cultivation and Utilization of Medicinal and Aromatic Plants. Regional Research Laboratory (CSIR); Jammu-Tawi: India:; 1982; p. 314-317.
- 3. Reynolds T. The compounds in Aloe leaf exudates: a review Botanical Journal of the Linnean Society.1985; 90:157-177.
- 4. Mathews CK, Van Holde KE. Biochemistry: The Benjamin/Cummings Publishing Company Inc; 1990.
- 5. Serfontein W. New nutrition Transform your life. Tafelberg Publishers; Cape Town: 2001.
- Ahmad I, Aqil F. *In vitro* efficacy of bioactive extracts of 15 medicinal plants against ESbLproducing multidrug-resistant enteric bacteria. Microbiol Res. 2007; 162: 264–275.
- Barbour EK, Al Sharif M, Sagherian VK, Habre AN, Talhouk RS, Talhouk SN. Screening of selected indigenous plants of Lebanon for antimicrobial activity. J Ethnopharmacol. 2004; 93:1-7
- Iwu MW, Duncan AR, Okunji CO. New antimicrobials of plant origin In: Perspectives on new Crops and new Uses eds. Janick J, ASHS Press Alexandria VA;1999. p. 457-462.
- 9. Butler MS. Natural products to drugs: natural product-derived compounds in clinical trials. Nat Prod Rep. 2008; 25: 475-516.
- Parkin DM. The global health burden of infectionassociated cancers in the year 2002. International Journal of Cancer. 2006; 118: 3030– 3044.
- 11. Triphathi KD. Textbook of Pharmacology and experimental therapeutics. India: Vallabh Prakashan; 1999.
- 12. Snow RW, Guerra CA, Noor AM. The global distribution of clinical episodes of *Plasmodium falciparum* malaria. Nature. 2005; 434: 214-217.
- 13. White NJ. Antimalarial drug resistance. Journal of Clinical Investigation. 2004; 113: 1084-1092.
- 14. Black JG. Microbiology: principles and explorations. 5th Ed. John Wiley & Sons Inc; New York: 2002.
- 15. Ozkan M, Dweik RA. Upper Respiratory Infection. Emedicine. 2004.
- 16. Dutta A, Sukul NC. Filaricidal properties of a wild herb *Andrographis paniculata.* J Helminthol.1982; 56: 81-84
- 17. Gallo RC. Human retroviruses after 20 years: A perspective from the past and prospects for their future control. Immunol Rev. 2002; 185: 236–265.
- Rasmussen HB, Perron H, Clausen J. Do endogenous retroviruses have etiological implications in inflammatory and degenerative nervous system diseases. Acta Neurol Scand. 1993; 88: 190–198.
- 19. Clausen J. Endogenous retroviruses and MS: using ERVs as disease markers; Int MS J. 2003; 10: 22–28.

- 20. Mishra SK, Sangwan NS, Sangwan RS. *Andrographis paniculata* (Kalmegh): A review. Pharmacog Rev. 2007; 1: 283-289.
- 21. Kanokwan J, Nobuo N. Pharmacological aspects of *Andrographis paniculata* on health and its major diterpenoid constitute andrographolide.J of Health Sci. 2008; 54: 370-381.
- 22. Pholphana N, Rangkadilok N, Thongnest S, Ruchirawat S, Ruchirawat M, Satayavivad J. Determination and variation of three active diterpenoids in *Andrographis paniculata* (Burm f) Nees. Phytochem Anal. 2004; 15: 365-371.
- 23. Ghosh Kumar B, Datta Kumar A, Mandal A, Kumari Priyanka D, Halder S. An overview on *Andrographis paniculata* (burm f) Nees. IJRAP. 2012; 3.
- 24. Medicinal plants in Viet Nam Manila World Health Organization (WHO Regional Publications 1990). Western Pacific: 3.
- 25. Standard of ASEAN herbal medicine. Jakarta ASEAN Countries. 1993; 1
- 26. Thai herbal pharmacopoeia. Bangkok Prachachon Co.1995;1.
- 27. Pharmacopoeia of the People's Republic of China. Beijing Chemical Industry Press; 1997; 1ed.
- 28. Bhardwaj A, Khatri P, Soni ML, Ali DJ. Potent herbal hepatoprotective drugs: a review. J Adv Sci Res. 2011; 2: 15-20.
- Bajaj YPS, Furmanowa M, Olszowska O. Biotechnology of the micropropagation of medicinal and aromatic plants In: Bajaj YPS, ed. Biotech Agri Forest 4 Medicinal and Aromatic Plants. Springer; Berlin, Germany: 1988. p.60-103.
- Pattnaik SK, Chand PK. *In vitro* propagation of the medicinal herbs *Ocimum americanum* (hoary basil) and *Ocimum sanctum* (holy basil). Plant Cell Rep. 1996; 15: 846-850.
- Purohit SD, Dave A, Kukda D. Micropropagation of safed musli (*Chlorophytum borivilianum*) a rare medicinal herb. Plant Cell Tiss Org Cult.1994; 39:93.
- 32. Rout GR. Direct plant regeneration from leaf explants of *Plambago* species and its genetic fidelity through RAPD markers. Annal Appl Biol. 2002;140: 305–313.
- Purkayastha J, Sugla T, Paul A, Solleti S, Sahoo L. Rapid *in vitro* multiplication and plant regeneration from nodal explants of *Andrographis paniculata*: A valuable medicinal plant. *In Vitro* Cell Dev Biol-Plant. 2008; 44: 442-447
- 34. Kataky A, Handique PJ. Micropropagation and screening of antioxidant potential of *Andrographis paniculata* (Burm f) Nees. Journal of Hill Agriculture. 2010a; 1:15-20.
- 35. Faisal M, Ahmad N, Anis M. Shoot multiplication in *Rauvolfia tetraphylla*L. using thidiazuron. Plant Cell Tiss Org Cult. 2005; 80: 187-190.
- Vijaya D, Padmadevi SN, Vasandha S, Meerabhai RS, Chellapandi P. Effect of vermicomposted coirpith on the growth of

Andrographis paniculata. J Org Syst. 2008;3: 51-56.

- 37. Samantaray S, Rout GR, Das P. Heavy metal and nutrient concentration in soil and plants growing on a metalliferous chromite. Minespoil Enviro Tech. 2001;22: 1147-54
- Kasetklangklung. Andrographis paniculata Thai medicinal plant. J Transfer Tech Agri. 1996; 2: 6-17
- 39. Zhou, Z. Cultivation of *Andrographis paniculata*. Chung Yao Tung Pao.1987;12: 15-18.
- 40. Muniramappa RP, Farooqi AA, Gowda HGR, Maricapu S. Influence of macronutrients on yield and active principle content in Kalmegh. J Med Arom Plant Sci.1997;19: 1039-1042.
- 41. Gupta V, Srivastava VK. Evaluation studies on Kalmegh (Andrographis *paniculata* Nees). Ind J Pl Genet Resour. 1995; 8:141-143.
- 42. Anonymous. 2000. The BIOME News, Vol 1. Arora JR, Renu Swarup, Gupta SV. Dept of Biotechnology Ministry of Science and Technology, Govt of India.
- 43. Lattoo SK, Khan S, Dhar AK, Choudhary DK, Gupta KK. Genetic and mechanism of induced male sterility in *Andrographis paniculata* (Burm f) Nees and its significance. Curr Sci. 2006; 9: 515-519.
- 44. Sharma A, Lai K, Handa SS. Standardization of Indian crude drug kalmegh by high performance liquid chromatographic determination of andrographolide. Phytochem Anal.1992; 3: 129-131.
- 45. Rastogi RP, Mehrotra BN. Compendium of Indian Medicinal Plants. Central Drug Research Institute, Lucknow: 1993; 52.
- Pholphana N, Rangkadilok N, Thongnest S, Ruchirawat S, Ruchirawat M, Satayavivad J. Determination and Variation of three active diterpenoids in *Andrographis paniculata* (Burmf) Nees. Phytochem Anal. 2004; 15: 365-371.
- 47. Pramanick S, Banerjee S, Achari B, Das B, Sen AK. Andrographolide and isoandrographolide minor diterpenoids from *Andrographis paniculata* Structure and X-ray crystallographic analysis. J Nat Prod. 2006; 69: 403-405.
- 48. Chen W, Liang X. Deoxyandrographolide 19 β-Dglucoside from the leaves of *Andrographis paniculata*. Planta Med.1982;15: 245-246.
- 49. Dastur JF. "Medicinal plants of India and Pakistan" Meyer Books: 1959.
- 50. Saxena S, Jain DC, Bhakuni RS, Sharma RP. Chemistry and pharmacology of Andrographis species. Indian Drugs.1998;35: 458-467.
- 51. Burgos RA, Loyola M, Hidalgo MA, Labranche TP, Hancke JL. Effects of 14deoxyandrographolide on calcium mediated rat uterine smooth muscle vontractility.Phytother Res. 2003;17: 1011-1015.
- 52. Zhang CY, Tan BK. Mechanism of cardiovascular acrivity of *Andrographis paniculata*in the anaesthetized rat.J Ethnopharmacol.1997;56: 97-101.
- 53. Zhang CY, Tan BK. Vasorelaxation of rat thoracic aorta caused by 14- deoxyandrographolide.Clin Exp Pharmacol Physiol.1998;25:424-429.

- 54. Kapil A, Koul IB, Banerjee SK, Gupta BD. Antihepatotoxic effects of major diterpenoid constituents of *Andrographis paniculata*.Biochem Pharmacol.1993;46:182-185.
- 55. Batkhuu J, Hattori K, Takano F, Fushiya S, Oshiman KI, Fujimiya Y. Suppression of NO production in activated macrophages in vitro and ex vivo by neoandrographolide isolated from *Andrographis paniculata*.Biol Pharm Bull. 2002;25:1169-1174
- Liu J, Wang ZT, Ji LL, Ge BX. Inhibitory effects of neoandrographolide on nitric oxide and prostaglandin E2 production in LPS-stimulated murine macrophage.Mol Cell Biochem.2007;298:49-57.
- 57. Yoopan N, Thisoda P, Rangkadilok N, Sahasitiwat S, Pholphana N, Ruchirawat S. Cardio vascular effects of 14-deoxy-1112didehydroandrographolide and *Andrographispaniculata* extracts.Planta Med.2007;73:503-511.
- 58. Zhang CY. Tan BK. Effects of 14deoxyandrographolide and 14-deoxy-1112didehydroandrographolide on nitric oxide production in cultured human endothelial cells.Phytother Res.1999;13:157-159.
- 59. Geethangili M, Rao YK, Fang SH, Tzeng YM. Cytotoxic constituents from *Andrographis paniculata*induce cell cycle arrest in Jurkat cells.Phytother Res.2008;22: 1336-1341.
- 60. Chao WW, Kuo YH, Lin BF. Anti-inflammatory Activity of New Compounds from *Andrographis paniculata*by NF-κB Trans-Activation inhibition.J Agric Food Chem.2010;58: 2505-2512.
- 61. Liu J, Wang ZT, Ge BX. Andrograpanin isolated from *Andrographis paniculata* exhibits antiinflammatory property in lipopolysaccharide induced macrophage cells through down regulating the p38 MAPKs signaling pathways.Int Immunopharmacol.2008;8: 951-958.
- Ji LL, Wang Z, Dong F, Zhang WB, Wang ZT. Andrograpanin a compound isolated from antiinflammatory traditional Chinese medicine *Andrographis paniculata* enhances chemokine SDF-1α-induced leukocytes chemotaxis.J Cell Biochem.2005;95: 970-978.
- Matsuda T, Kuroyanagi M, Sugiyama S, Umehara K, Ueno A, Nishi K. Cell differentiation inducing diterpenes from *Andrographis paniculata*Nees.Chem Pharm Bull. 1994;42: 1216-1225.
- 64. Chen L, Zhu H, Wang R, Zhou K, Jing Y, Qiu F.Ent-Labdane diterpenoid lactone stereoisomers from *Andrographis paniculata.*J Nat Prod.2008;71:852-855.
- 65. Jada SR, Suseno GS, Matthews C, Hamzah AS, Lajis NH, Saad MS, Stevens MFG, Stanslas J. Semisynthesis and in vitro anticancer activities of andrographolide
 - analogues.Phytochemistry.2007;68:904-912.
- 66. Ojha SK, Nandave M, Kumari S, Ary DS. Antioxidant activity of *Andrographis paniculata* in ischemic myocardium of rats. Global J Pharmacol.2009;3:154-157.

- 67. Shen YC, Chen CF, Chiou WF. Andrographolide prevents oxygen radical production by human neutrophils: possible mechanism(s) involved in its anti-inflammatory effect. Br J Pharmacol.2002;135:399-406.
- 68. Dandu AM, Inamdar NM. Evaluation of beneficial effects of antioxidant properties of aqueous leaf extract of *Andrographis paniculata* in STZ-induced diabetes. Pak J Pharm Sci.2009;22:49-52.
- 69. Yu BC, Hung CR, Chen WC, Cheng JT. Antihyperglycemic effect of andrographolide in streptozotocin-induced diabetic rats. Planta Med.2003;69:1075-1079.
- 70. Lopez A, Mathers C, Ezzati M, Jamison D, Murray C. Global and regional burden of disease and risk factors 2001: systematic analysis of population health data. Lancet.2006;367:1747-1757.
- 71. Gupta S, Choudhry MA, Yadava JNS. Antidiarrioeal activity diterpenes of *Andrographis paniculata* (Kal-Megh) agent *Escherichia coli* enterotoxin in vivo models. International Journal of Crude Drug Research.1990;283-284.
- 72. Thanangkul P, Chaichantipyuth C. Clinical studies of *Andrographis paniculata* on diarrhoea and dysentery. Journal of Ramatipodee Medical Sciences of Thailand. 1985;8: 57-62.
- 73. Pleumjai T, Sithisomwongse N. 1990 Antimicrobial activity of *Andrographis paniculata* Nees. Symposium on *Andrographis paniculata*, National Institute of Health; Nonthaburi Thailand.
- 74. Tomoko N, Takashi A, Hiromu T,Yuka I, Hiroko M. Antibacterial activity of extracts prepared from tropical and subtropical plants on methicillin resistant *Staphylococcus aureus*. J Health Sci.2002;48:273-276.
- 75. Mishra PK, Singh RK, Gupta A, Chaturvedi A, Pandey R. Antibacterial activity of *Andrographis paniculata* (Burm F)Wall ex Nees leaves against clinical pathogens. JPR.2013;**7**:459-462.
- 76. Premanath R, Devi NL. Antibacterial antifungal and antioxidant activities of *Andrographis paniculata* Nees leaves. International Journal of Pharmaceutical Sciences.2011;2:2091-2099.
- 77. Deepak S, Pawar A, Shinde P. Study of antioxidant and antimicrobial activities of *Andrographis paniculata*. Asian Journal of Plant Science and Research.2014;4:31-41.
- 78. Shirisha K, Mastan M. *Andrographis paniculata* and its bioactive phytochemical constituents for oxidative damage: a systematic review. Pharmacophore.2013;4:212-229.
- 79. Manjusha G, Rajathi K, Mini Alphonse JK, Meera K. Antioxidant potential and antimicrobial activity of *Andrographis paniculata* and *Tinospora cordifolia* against pathogenic organisms. Journal of Pharmacy Research.2011;4:452.
- 80. Bobbarala V, Koteswara Rao P, Srinivasa Rao G, Aryamithra D. Bioactivity of *Andrographis paniculata* against selected phytopathogens. Journal of Pharmacy Research.2009.
- 81. Singha PK, Roy S, Dey S. Antimicrobial activity of Andrographis paniculata. Fitoterapia.2003;74:692-694.

- Kondo S, Chatuphonprasert W, Jaruchotikamol A, Sakuma T, Nemoto N. Cellular glutathione content modulates the effect of andrographolide on l²-naphthoflavone-induced CYP1A1 mRNA expression in mouse hepatocytes. Toxicology.2011;280:18-23.
- 83. Sirisha Mulukuri NVL, Mondal NB, Raghu Prasad M, Renuka S, Ramakrishna K. Isolation of Diterpenoid Lactones from the Leaves of *Andrographis paniculata* and Its Anticancer Activity. International Journal of Pharmacognosy and Phytochemical Research.2011;3:39-42.
- 84. Arsenault PR, Wobbe KK, Weathers PJ. Recent advances in artemisinin production through heterologous expression. Curr Med Chem.2008;15:2886-2896.
- 85. Mishra K, Dash AP, Swain BK, Dey N. Antimalarial activities of *Andrographis paniculata* and *Hedyotis corymbosa* extracts and their combination with curcumin. Malar J.2009;8:26.
- Caceres DD, Hancke JL, Burgos RA, Wikman GK. Prevention of common colds with *Andrographis paniculata* dried extract: A Pilot double blind trial. Phytomedicine.1997;4: 101-104.
- 87. The promotion and development of traditional medicine: report of a WHO meeting World Health Organ. Tech Rep Ser.1978;1-41.
- Xia YF, Ye BQ, Li YD, Wang JG, He XJ. Andrographolide attenuates inflammation by inhibition of NF-kappa B activation through covalent modification of reduced cysteine 62 of p50. J Immunol.2004;173:4207-4217.
- Hidalg MA, Romero A, Figueroa J, Cortés P, Concha II. Andrographolide interferes with binding of nuclear factor-kappaB to DNA in HL-60-derived neutrophilic cells. Br J Pharmacol.2005;144:680-686.
- 90. DMPRD. Division of Medical Plants Research and Development. Department of Medical Science, Ministry of Public Health: Hand book of medicinal plant for primary public health; Text and Journal Corporation Co Ltd. Bangkok:1990;53 p
- 91. Amroyan E, Gabrielian E, Panossian A, Wikman G, Wagner H. Inhibitory effect of andrographolide from *Andrographis paniculata* on PAF-induced platelet aggregation. Phytomedicine.1999;6:27-31.
- Wiart C, Kumar K, Yusof MY, Hamimah H, Fauzi ZM. Antiviral properties of ent-labdene diterpenes of *Andrographis paniculata* Nees inhibitors of herpes simplex virus type 1. Phytother Res.2005;19:1069-1070.
- 93. Dua VK, Verma G, Dash AP. In vitro antiprotozoal activity of some xanthones isolated from the roots of *Andrographis paniculata*. Phytother Res.2009;23:126-128.
- 94. Radha R, Sermakkani M, Thangapandian V. Evaluation of phytochemical and antimicrobial activity of *Andrographis paniculata* Nees (Acanthaceae) aerial parts. IJPLS.2011;2:562-567.
- 95. Aniel Kumar O, Mutyala Naidu L, Raja Rao KG. In vitro antibacterial activity in the extracts of *Andrographis paniculata* Burm. International

Journal of Pharm Tech Research.2010;2:1383-1385.

- 96. Goel SR, Madan VK, Verma KK, Nandal SN. Nematicidal activity of various medicinal and aromatic plants under in vitro conditions. Indian Journal of Nematology.2009;39: 218-220.
- 97. Tripathi AK, Prajapati V, Jain DC, Saxena S. 1999 Antifeedant oviposition deterrent and growthinhibitory activity of *Andrographis paniculata* against *Spilarctia oblique*.Int J Trop Insect Sci. 1999;19:211-216.
- Govindarajan M. Evaluation of Andrographis paniculata Burmf(Family:Acanthaceae) extracts against Culex quinquefasciatus (Say) and Aedes aegypti (Linn) (Diptera:Culicidae). Asian Pac J Trop Med,2011;4:176-181.
- 99. Choudhary BR, Poddar MK. 1984 Andrographolide and kalmegh (*Andrographis paniculata*) extract in vivo and in vitro effect on lipid peroxidation. Meth Find Exp Clin Pharmacol. 1984;6:481-485.
- 100. Trivedi N, Rawal UM. Hepatoprotective and toxicological evaluation of *Andrographis paniculata* on severe liver damage. Indian J Pharmacol.2000;32:288- 293.
- 101. Singh RP, Banerjee S, Rao AR. Modulatory influence of *Andrographis paniculata* on mouse hepatic and extrahepatic carcinogen metabolizing enzymes and antioxidant status. Phytother Res.2001;15:382-290.
- 102. Jarukamjorn K, Don-in K, Makejaruskul C, Laha T, Daodee S, Pearaksa P, Sripanidkulchai B. Impact of Andrographis paniculata crude extract on mouse hepatic cytochrome P450 enzymes.J Ethnopharmacol.2006;105:464-467.
- 103. Tan BK, Zhang C, Kuroyangi M. Cardiovascular activity of 14-deoxy-11 12didehydroandrographolide in the anaesthetized rat and isolated right atria. Pharmacol Res.1998;38:413–417.
- 104. Naidu S, Asmawi M, Amirin S. Vasorelaxant effect of chloroform extract of *Andrographis paniculata* on in-vitro rat thoracic aorta. 2007;12 p
- 105. Karaki H, Nakagawa H, Urakawa N. Comparative effects of Verapamil and sodium nitroprusside on contraction and 45Ca uptake in the smooth muscle of rabbit aorta rat aorta and guinea-pig taenia coli. Br J Pharmacol.1984b;81:393-400.
- 106. Murugan K, Selvanayaki K, Kalyanasundaram VB, Sohaibani SA. Nanotechnological approach for exploring the antibiofilm a potential of an ethanomedicinal herb *Andrographis paniculata* for controlling lung infection causing *Pseudomonas aeruginosa*. Digest J Nanomaterials Biostructures.2013b;8:117-126.
- 107. Bosgelmez-Tinaz G, Ulusoy S, Ugur A, Ceylan O. Inhibition of quorum sensing–regulated behaviors by *Scorzonera sandrasica*. Curr Microbiol.2007;55:114–118.
- 108. Sohaibani SA, Murugan K. Anti-biofilm activity of *Salvodora persica* on cariogenic isolates of *Streptococcus mutans*: in-vitro and molecular docking studies. Biofouling.2012;28:29-38.
- 109. Ertl P. Database of bioactive ring systems with calculated properties and its use in bioisosteric

design and scaffold hopping. Bioorg Med Chem.2012;20:5436–5442.

- 110. Akbar S."*Andrographis paniculata*: a review of pharmacological activities and clinical effects."Alternative Medicine Review. 2011;16 (1) p. 66–77.
- 111. Kabir MH,Hasan N,Rahman MM. "A survey of medicinal plants used by the Deb barma clan of the Tripura tribe of Moulvibazar district Bangladesh." Journal of Ethnobiology and Ethnomedicine.2014;10(1) article 19.
- 112. Niranjan A, Tewari SK, Lehri A. "Biological activities of Kalmegh (*Andrographis paniculata*Nees) and its active principles-A review." Indian Journal of Natural Products and Resources.2010;1(2)125–135.
- 113. Wu Z, Raven PH, Hong DY. Garden Flora of China: Cucurbitaceae Through Valerianaceae with Annonaceae and Berberidaceae. Science Press; Beijing, China: 1996.
- Benoy GK, Animesh DK, Aninda M, Priyanka DK, Sandip H. "An overview on Andrographis paniculata (burm F) Nees." International Journal of Research in Ayurveda and Pharmacy.2012; 3(6)752–760.
- 115. Jarukamjorn K, Nemoto N. "Pharmacological aspects of *Andrographis paniculata* on health and its major diterpenoid constituent andrographolide." Journal of Health Science. 2008;54(4)370–381.
- 116. Valdiani A, Kadir MA, Tan SG, Talei D, Abdullah MP, Nikzad S. "Nain-e havandi *Andrographis paniculata* present yesterday absent today: a plenary review on underutilized herb of Iran's pharmaceutical plants." Molecular Biology Reports. 2012;39(5)5409–5424.
- 117. Sharma M, Sharma R. "Identification purification and quantification of andrographolide from*Andrographis paniculata* (burm F) Nees by HPTLC at different stages of life cycle of crop." Journal of Current Chememical and Pharmaceutical Sciences.2013;3(1)23–32.
- 118. Boopathi C."Andrographis spp: a source of bitter compounds for medicinal use." Ancient Science of Life.2000;19(3-4)164–168.
- 119. Anju D, Jugnu G, Kavitha S, Arjun N, Sandeep D. "A review on medicinal prospective of *Andrographis paniculata* Nees." Journal of Pharmaceutical and Scientific Innovation.2012;1(1)1–4.
- 120. Tholl D, Lee S. Terpene specialized metabolism in *Arabidopsis* thaliana. Arabidopsis Book. 2011;9:0143-44.
- 121. De-Eknamkul W, Potduang B. Biosynthesis of beta-sitosterol and stigmasterol in Croton sublyratus proceeds via a mixed origin of isoprene units. Phytochemistry. 2003;62:389– 398.
- 122. Paetzold H, Garms S, Bartram S, Wieczorek J, Uros-Gracia EM, Rodriguez-Concepcion M. The isogene 1-deoxy-D-xylulose 5-phosphate synthase 2 controls isoprenoid profiles precursor pathway allocation and density of tomato trichomes. Mol Plant. 2010;3:904–916.

123. Schramek N, Wang H, Romisch-Margl W, Keil B, Radykewicz T, Winzenhorlein B. Artemisinin biosynthesis in growing plants of Artemisia annua A 13CO2

study. Phytochemistry. 2010;71:179–187.

- 124. Srivastava N, Akhila A. Biosynthesis of andrographolide in *Andrographis paniculata*. Phytochemistry.2010;71:1298–1304.
- 125. Choudhary S, Gaur R, Gupta S. EST-derived genic molecular markers: development and utilization for generating an advanced transcript map of chickpea. Theor Appl Genet. 2012;124:1449–1462.
- 126. Hedden P, Kamiya Y. Gibberellin biosynthesis enzymes genes and their regulation. Annu Rev Plant Physiol Plant Mol Biol. 1997;48:431–460.
- 127. Hartweck LM. Gibberellin signaling. Planta. 2008;229:1–13.
- 128. Cyr A, Wilderman PR, Determan M, Peters RJ. A modular approach for facile biosynthesis of labdane-related diterpenes. J Am Chem Soc. 2007;129:6684–6685.
- 129. Peters RJ. Two rings in them all the labdanerelated diterpenoids. Nat Prod Rep. 2010;27:1521–1530.
- 130. Otomo K, Kenmoku H, Oikawa H, Konig WA, Toshima H, Mitsuhashi W. Biological functions ofent- and syn-copalyl diphosphate synthases in rice key enzymes for the branch point of gibberellin and phytoalexin biosynthesis. Plant J. 2004;39:886–893.
- 131. Prisic S, Xu M, Wilderman PR, Peters RJ. Rice contains two disparate ent-copalyl diphosphate synthases with distinct metabolic functions. Plant Physiol. 2004;136:4228–4236.
- 132. Wu Y, Zhou K, Toyomasu T, Sugawara C, Oku M, Abe S. Functional characterization of wheat copalyl diphosphate synthases sheds light on the early evolution of labdane-related diterpenoid metabolism in the cereals. Phytochemistry.2012;84:40–46.
- Shimura K, Okada A, Okada K, Jikumaru Y, Ko KW, Toyomasu T. Identification of a biosynthetic gene cluster in rice for momilactones. J Biol Chem. 2007;282:34013–34018.
- 134. WHO Technical Report Series: Geneva Trace Elements in Human Nutrition and Health. 1996;119-205.
- 135. Diaz-Gomez NM, Domenech E, Barroso F, Castells S, Cortabarria C, Jimenz A. The Effect of Zinc Supplementation on Linear Growth Body Composition and Growth Factors. Preterm Infants Pediatrics. 2003;111(5):1002-1009.

- 136. Neetu Sharma, Rajendra Prasad, Kala, Bhawana Kweera. Comparative Study of the Elemental Composition of Swertia Chirayita from Two Different Sites of Garhwal in Uttrakhand; Int J Chem Sci. 2013;11(4):1713-1720.
- 137. Farooqui AA, Sreeramu BS. Kalmegh in: Cultivation of aromatic and medicinal crop. Universities press Ltd; Hyderabad:2001;151-156.
- 138. Samy RP, Thwin MM, Gopalakrishnakone P, Ignacimuthu S. Ethanobotanical survey of for the treatment of snakebites in southern part of Tamil Nadu.J Ethnopharmacol. 2008;115:302-312.
- Martin KP.Plant regeneration protocol of medicinally important Andrographis paniculata (burm f) Wallich ex nees via somatic embryogenesis. In vitro Cell Dev Biol Plant.2004;40:204-209.
- 140. Akbar S.*Andrographis paniculata*: A review of pharmacological activities and clinical effects. Int J health sci. 16:66-77.
- 141. Burgos RA, Seguel K, Perez M, Meneses A, Ortega M. Andrographolide inhibits IFN-gamma and IL-2 cytokine production and protects against cell apoptosis. Planta Med.2005;71: 429-434.
- 142. Bharat Kumar, Divya Topal. Effect of Organic Fertilizers on the Growth of Shoot of Kalmegh (*Andrographis Paniculata*). International Journal of Novel Research in Life Sciences.2015;2:9-11.
- 143. Verma N, Vinayak M. Antioxidant action of *Andrographis paniculata* on lymphomaMol Biol Rep. 2008;35(4):535-540.
- 144. Radhika A, Annapurna S, Nageswara Rao. Immunostimulant, cerebroprotective & nootropic activities of *Andrographis paniculata* leaves extract in normal & type 2 diabetic rats. Indian P J Med Res. 2012;135:636-641.
- 145. Wibudi A, Kiranadi B, Manalu W. Traditional plant, *Andrographis paniculata* (Sambiloto), exhibits insulin-releasing actions in vitro. Acta Med Indones. 2008; 40:63-68
- Dua VK, Ojha VP, Roy R. Antimalarial activity of some xanthones isolated from the roots of *Andrographis paniculata*. J Ethnopharmacol. 2004;95:247-251.
- 147. Deng WL, Nie RJ, Liu JY (1982): Comparison of pharmacological effect of four andrographolides. Chinese Pharmaceutical Bulletin 17: 195-198
- 148. Stephen H, Comac L. 2000. Miracle herbs: How herbs combine with modern medicine to treat cancer, heart disease, AIDS and more, Kensington publishing corporation, New York.